# Performance Comparison of 15 Transport Ventilators 

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#### Abstract

BACKGROUND: Numerous mechanical ventilators are designed and marketed for use in patient transport. The complexity of these ventilators differs considerably, but very few data exist to compare their operational capabilities. METHODS: Using bench and animal models, we studied 15 currently available transport ventilators with regard to their physical characteristics, gas consumption (duration of an E-size oxygen cylinder), battery life, ease of use, need for compressed gas, ability to deliver set ventilation parameters to a test lung under 3 test conditions, and ability to maintain ventilation and oxygenation in normal and lung-injured sheep. RESULTS: Most of the ventilators tested were relatively simple to operate and had clearly marked controls. Oxygen cylinder duration ranged from 30 min to 77 min . Battery life ranged from 70 min to 8 hours. All except 3 of the ventilators were capable of providing various $\mathrm{F}_{\mathrm{IO}_{2}}$ values. Ten of the ventilators had high-pressure and patient-disconnect alarms. Only 6 of the ventilators were able to deliver all settings as specifically set on the ventilator during the bench evaluation. Only 4 of the ventilators were capable of maintaining ventilation, oxygenation, and hemodynamics in both the normal and the lung-injured sheep. CONCLUSIONS: Only 2 of the ventilators met all the trial targets in all the bench and animal tests. With many of the ventilators, certain of the set ventilation parameters were inaccurate (differed by $\mathbf{> 1 0 \%}$ from the values from a cardiopulmonary monitor). The physical characteristics and high gas consumption of some of these ventilators may render them less desirable for patient transport. Key words: transport, mechanical ventilation, ventilator, positive end-expiratory pressure, PEEP, fraction of inspired oxygen, $F_{I O_{2}}$. [Respir Care 2007;52(6): 740-751. © 2007 Daedalus Enterprises]


## Introduction

Patients who require ventilatory support are frequently transported from one hospital location to another. Portable

[^0]transport ventilators are also required in ambulances and in forward military positions. In addition, the threat of bioterrorism requires health care systems to be able and rapidly with very little notice to accept and ventilate large numbers of patients.

To provide ventilatory support under the above-defined conditions requires that transport ventilators be appropriately designed, though this does not mean they must be equivalent to intensive care unit ventilators. ${ }^{1}$ Transport ventilators must incorporate certain characteristics to be of use in the above-defined settings. ${ }^{1-3}$ First, they must be able to ventilate patients with healthy or acutely or chronically injured lungs. Second, they must be portable and easy to operate. Third, they must be able to deliver a high fraction of inspired oxygen $\left(\mathrm{F}_{\mathrm{IO}_{2}}\right)$. Fourth, in forward military positions they must be able to operate on an internal battery for a long period, and without compressed gas.

[^1]Table 1. Comparison of Evaluated Ventilators by Power Source, Physical Dimensions, Modes, $\mathrm{F}_{\mathrm{IO}_{2}}$, and PEEP

| Ventilator | Approved as <br> Transport <br> Ventilator | Power Requirement | Size |  |  | Weight (kg) | Available Modes | $\mathrm{F}_{\mathrm{IO}_{2}}$ (range or available values) | PEEP <br> Available? <br> (range <br> cm $\mathrm{H}_{2} \mathrm{O}$ ) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  | Height (cm) | Width (cm) | Depth <br> (cm) |  |  |  |  |
| Sophisticated |  |  |  |  |  |  |  |  |  |
| Univent Eagle 754 | Yes | Electricity | 29 | 23 | 11 | 4.5 | A/C, SIMV | 0.21-1.0 | Yes, 0-20 |
| VersaMed iVent | Yes | Electricity | 33 | 24 | 26 | 10 | $\begin{aligned} & \text { A/C (VP), SIMV (VP), } \\ & \text { PSV, CPAP } \end{aligned}$ | 0.21-1.0 | Yes, 0-20 |
| Newport HT50 | Yes | Electricity | 26 | 27 | 20 | 6.8 | $\begin{aligned} & \text { A/C (VP), SIMV (VP), } \\ & \text { PSV, CPAP } \end{aligned}$ | 0.21-1.0 | Yes, 0-30 |
| Pulmonetic Systems LTV 1000 | Yes | Electricity | 8 | 25 | 30 | 6.1 | $\begin{aligned} & \text { A/C (VP), SIMV (VP), } \\ & \text { PSV, CPAP } \end{aligned}$ | 0.21-1.0 | Yes, 0-30 |
| Simple |  |  |  |  |  |  |  |  |  |
| Oceanic Medical Products Magellan | Yes | Gas | 12.7 | 17.8 | 10.2 | 2.1 | CMV, IMV | 1.0 | No |
| Bio-Med Devices IC2A | Yes | Gas | 26 | 16 | 9 | 4.1 | A/C, SIMV | 1.0 | Yes, 0-30 |
| Pneupac Parapac Medic | Yes | Gas | 9.2 | 22 | 16.2 | 2.4 | CMV, SIMV | 0.5, 1.0 | No |
| Pneupac Parapac Transport 200D | Yes | Gas | 9.2 | 22 | 16.2 | 3.1 | CMV, SIMV | 0.5, 1.0 | No |
| Life Support Products Auto Vent 2000 | Yes | Gas | 15 | 4.5 | 9 | 0.68 | CMV | 1.0 | No |
| Carevent ATV+ | Yes | Gas | 23.5 | 11.1 | 16.2 | 4.1 | CMV, IMV | 0.6,1.0 | Yes, 0-20 |
| Vortran RespirTech Pro | No | Gas | 16.76 | 6.35 | 8.38 | 0.165 | CMV, IMV | 1.0 | No |
| Percussionaire TXP | Yes | Gas | 10.6 | 10.6 | 16.5 | 0.68 | CMV, IMV | 0.5 | No |
| Bio-Med Devices Crossvent 3 | Yes | Gas + electricity | 22.9 | 28 | 12.7 | 4.32 | $\begin{aligned} & \text { A/C (VP), SIMV (VP), } \\ & \text { PSV, CPAP } \end{aligned}$ | 0.5, 1.0 | No |
| Bird Avian | Yes | Gas + electricity | 25 | 30 | 12.7 | 4.5 | A/C, SIMV | 1.0 | No |
| Pneupac Compac 200 | Yes | Gas or electricity | 36 | 21 | 21 | 8.5 | CMV, IMV | 0.45, 1.0 | No |

[^2]Fifth, they should be able to provide both assisted and controlled ventilation. Sixth, they must incorporate alarms that identify catastrophic conditions.

Previous evaluations of transport ventilators included only up to 8 ventilators. ${ }^{4-10}$ Many of the ventilators previously evaluated have since been modified by the manufacturers, and new ventilators have entered the market. We present an evaluation of 15 transport ventilators for use during intrahospital or ambulance transport and in forward military positions. The goals of this study were (1) to determine if these transport ventilators could ventilate both healthy and injured lungs, and deliver tidal volumes $\left(\mathrm{V}_{\mathrm{T}}\right)$ and respiratory rates ( RR ) as specifically set, and (2) to identify which ventilators would be most appropriate in which transport settings.

## Methods

Table 1 shows power requirements, physical dimen-
sions, available modes, available $\mathrm{F}_{\mathrm{IO}_{2}}$ ranges and settings, and positive end-expiratory pressure (PEEP) range for the 15 tested ventilators. All 15 ventilators were provided by their respective manufacturers specifically for this evaluation. All 15 ventilators are approved by the U.S. Food and Drug Administration for use in transport, except the Vortran RespirTech Pro, which is marketed as a resuscitator.

## Bench Protocol

We evaluated gas consumption, battery life, ease of use, physical characteristics, need for compressed gas, and the ability to deliver set ventilation parameters under 3 different test conditions. Gas consumption was defined as the amount of time the ventilator could function on one full E-size oxygen cylinder (capacity 660 L of oxygen), with the ventilator set to deliver a $\mathrm{V}_{\mathrm{T}}$ of $1,000 \mathrm{~mL}$ at an RR of 10 breaths/min and an $\mathrm{F}_{\mathrm{IO}_{2}}$ of 1.0. Battery life was defined
as the amount of time the ventilator could function on a fully charged battery with the ventilator set to deliver a $\mathrm{V}_{\mathrm{T}}$ of $1,000 \mathrm{~mL}$ at an RR of 10 breaths $/ \mathrm{min}$ and an $\mathrm{F}_{\mathrm{IO}_{2}}$ of 0.21 .

A ventilator was considered easy to use if all the parameters were clearly labeled and easily set to deliver a precise variable (eg, $\mathrm{V}_{\mathrm{T}}$ or RR ). We assumed the manufacturer's published weight and dimensions to be accurate. Ability to ventilate without compressed gas was met if the ventilator could deliver the set minute volume $\left(\dot{V}_{\mathrm{E}}\right)$ under each of the test conditions without a compressed gas source or an external compressor.

The ability to deliver set parameters was evaluated with a test lung (Training and Test Lung, Michigan Instruments, Grand Rapids, Michigan) under 3 different test conditions: high resistance with normal compliance; normal resistance with normal compliance; and normal resistance with low compliance. High and normal resistance was achieved with resistors (Pneuflo Rp20 and Rp5, Michigan Instruments, Grand Rapids, Michigan). Normal and low compliance were set on the test lung $\left(0.05 \mathrm{~L} / \mathrm{cm} \mathrm{H}_{2} \mathrm{O}\right.$ and $0.02 \mathrm{~L} / \mathrm{cm} \mathrm{H}_{2} \mathrm{O}$, respectively). For each condition the tested ventilator was set to deliver a $\mathrm{V}_{\mathrm{T}}$ of 500 mL at 15 breaths $/ \mathrm{min}$ and 30 breaths $/ \mathrm{min}$, and a $\mathrm{V}_{\mathrm{T}}$ of 1 L at 10 breaths/min and 20 breaths $/ \mathrm{min}$.
$\mathrm{V}_{\mathrm{T}}, \mathrm{RR}$, peak inspiratory pressure (PIP), and positive endexpiratory pressure (PEEP) were measured and analyzed with a cardiopulmonary monitor (NICO, Respironics, Wallingford, Connecticut) and its software (Analysis Plus, Respironics, Wallingford, Connecticut). Ventilator performance was determined by comparing the set parameters to the measurements from the cardiopulmonary monitor.

Each ventilator was bench tested as follows. With resistance set at $20 \mathrm{~cm} \mathrm{H}_{2} \mathrm{O} / \mathrm{L} / \mathrm{s}$ and compliance set at 0.05 $\mathrm{L} / \mathrm{cm} \mathrm{H}_{2} \mathrm{O}$, the ventilator was connected to the test lung, and the cardiopulmonary monitor's flow sensor was placed between the ventilator circuit and the flow resistor. $\mathrm{V}_{\mathrm{T}}$ was initially set at $500 \mathrm{~mL}, \mathrm{RR}$ at 15 breaths $/ \mathrm{min}$, inspiratory time $\left(\mathrm{T}_{\mathrm{I}}\right)$ at 1.0 s (if setting the $\mathrm{T}_{\mathrm{I}}$ was possible on that ventilator), and PEEP at $5 \mathrm{~cm} \mathrm{H}_{2} \mathrm{O}$ (if PEEP was available on the ventilator). The $\mathrm{F}_{\mathrm{IO}_{2}}$ was set at the lowest available setting, which may have been 0.21 , air mix (entrainment), or 1.0 , depending on the ventilator's capabilities. Following a 10-breath stabilization period, we recorded PIP, mean airway pressure, PEEP, $\mathrm{V}_{\mathrm{T}}, \dot{\mathrm{V}}_{\mathrm{E}}, \mathrm{RR}$, and the pressure, flow, and volume graphics. After that data collection, the $R R$ was increased to 30 breaths $/ \mathrm{min}$ and the $T_{I}$ was decreased to 0.5 s . All other settings remained unchanged. Following another 10-breath stabilization period, we again recorded PIP, mean airway pressure, PEEP, $\mathrm{V}_{\mathrm{T}}, \dot{\mathrm{V}}_{\mathrm{E}}, R R$, and graphics. The $\mathrm{V}_{\mathrm{T}}$ was then increased to $1,000 \mathrm{~mL}, \mathrm{RR}$ was decreased to 10 breaths $/ \mathrm{min}$, and $\mathrm{T}_{\mathrm{I}}$ was increased to 1 s . Following another stabilization period and data collection, the RR was increased to 20 breaths $/ \mathrm{min}$, and sta-
bilization and data collection were repeated. All other settings remained unchanged. These settings were repeated for each of the compliance and resistance combinations. The cardiopulmonary monitor was interfaced with a laptop computer, on which the flow, volume, and pressure data were collected and analyzed. The set ventilation parameters, ventilator-displayed values, and cardiopulmonary-monitor-measured values were simultaneously recorded. All measurements during the bench assessment were at atmospheric-temperature-and-pressure-dry conditions.

## Laboratory Protocol

This protocol was approved by the animal care committee of Massachusetts General Hospital, Boston, Massachusetts.

Using $30-\mathrm{kg}$ sheep, we evaluated each ventilator's ability to ventilate both healthy and saline-lavage lung-injured sheep. In both settings we evaluated the ventilator's ability to maintain normal arterial blood gas values and cardiopulmonary hemodynamics. We studied 12 sheep: 6 with normal lungs and 6 with saline-lavage lung injury. Five ventilators were evaluated on each sheep (healthy and injured), and each group of 5 ventilators was studied on 2 healthy and 2 injured sheep. Three groups of 5 ventilators were randomly selected.

## Healthy Lung Evaluation

Each group of ventilators was randomly applied for a 60-min period to a healthy sheep. Initially, each ventilator was set at a $\mathrm{V}_{\mathrm{T}}$ of $9 \mathrm{~mL} / \mathrm{kg}$ and an RR of 20 breaths $/ \mathrm{min}$, with a $T_{I}$ or peak flow setting to maintain a $T_{I}$ of 1.0 s . If the device was capable of applying PEEP, PEEP of $5 \mathrm{~cm} \mathrm{H}_{2} \mathrm{O}$ was applied with $50 \%$ oxygen. The ventilator was attached to the animal's airway, followed by a $15-\mathrm{min}$ stabilization period. After stabilization we collected arterial and mixed venous blood samples, and measured systemic arterial pressure, pulmonary artery pressure, pulmonary capillary wedge pressure, and heart rate. Airway pressure and $\mathrm{V}_{\mathrm{T}}$ were measured at the endotracheal tube (ETT). Cardiac output was measured in triplicate, using the thermodilution technique. The ventilator was adjusted and oxygen added if needed to attempt to reach the target blood gas values $\left(\mathrm{P}_{\mathrm{aO}_{2}} 60-100 \mathrm{~mm} \mathrm{Hg}, \mathrm{P}_{\mathrm{aCO}_{2}} 30-\right.$ $50 \mathrm{~mm} \mathrm{Hg}, \mathrm{pH} 7.30-7.50$ ). Once we determined whether the targets could be met, the next ventilator was attached to the animal's airway for evaluation.

## Injured Lung Evaluation

During the lung-injury tests, the ventilator was initially set at a $\mathrm{V}_{\mathrm{T}}$ of $6 \mathrm{~mL} / \mathrm{kg}$, an RR of 30 breaths/min, PEEP of $15 \mathrm{~cm} \mathrm{H} \mathrm{H}_{2} \mathrm{O}$ (if available), and $\mathrm{F}_{\mathrm{IO}_{2}}$ of 0.50 . Again, blood
gases and hemodynamics were evaluated to determine if the target blood gas values $\left(\mathrm{P}_{\mathrm{aO}} 60-100 \mathrm{~mm} \mathrm{Hg}, \mathrm{P}_{\mathrm{aCO}}^{2}\right.$ $30-50 \mathrm{~mm} \mathrm{Hg}, \mathrm{pH} 7.30-7.50$ ) had been met, then the ventilator was adjusted as necessary to attempt to meet the targets. Once we determined whether the targets could be met, the next ventilator was attached to the animal's airway for evaluation.

## Instrumentation

We used 12 female Dorset sheep ( $21-31 \mathrm{~kg}$ ), each fasted for 24 hours. Orotracheal intubation, with an $8-\mathrm{mm}$ innerdiameter ETT, was performed during deep halothane anesthesia via mask. The external jugular vein was then cannulated, and an 8 French sheath introducer was inserted. After line placement, the anesthesia delivery was changed to intravenous only, with a loading dose of $10 \mathrm{mg} / \mathrm{kg}$ pentobarbital, $4 \mathrm{mg} / \mathrm{kg}$ ketamine, and $0.1 \mathrm{mg} / \mathrm{kg}$ pancuronium, followed by continuous infusion of pentobarbital ( $4 \mathrm{mg} / \mathrm{kg} / \mathrm{h}$ ), ketamine ( $8 \mathrm{mg} / \mathrm{kg} / \mathrm{h}$ ), and pancuronium ( $0.1 \mathrm{mg} / \mathrm{kg} / \mathrm{h}$ ) to provide surgical anesthesia with paralysis. After intubation, the basic ventilatory settings were volume control ventilation at a $\mathrm{V}_{\mathrm{T}}$ of $10 \mathrm{~mL} / \mathrm{kg}$, inspira-tory-expiratory ratio of $1: 2, \mathrm{~F}_{\mathrm{IO}_{2}}$ of 1.0 , and PEEP of $5 \mathrm{~cm} \mathrm{H}_{2} \mathrm{O}$, delivered by an intensive care ventilator ( 840 , Puritan Bennett, Carlsbad, California). RR was adjusted to achieve eucapnia ( $\mathrm{P}_{\mathrm{aCO}_{2}} 35-45 \mathrm{~cm} \mathrm{H} \mathrm{H}_{2} \mathrm{O}$ ).

An 18 -gauge catheter was then placed into the carotid artery for continuous measurement of arterial blood pressure and sampling of arterial blood gas values. Arterial and mixed venous blood samples were drawn for blood gas analysis. $\mathrm{P}_{\mathrm{O}_{2}}, \mathrm{P}_{\mathrm{CO}_{2}}, \mathrm{pH}$, oxyhemoglobin saturation, and hemoglobin content were assessed with a blood gas analyzer (282, Ciba Corning Diagnostics, Norwood, Massachusetts). Flow at the ETT was measured by a heated pneumotachometer (Hans Rudolph, Kansas City, Missouri) connected to a differential pressure transducer (MP- $45 \pm 2 \mathrm{~cm} \mathrm{H} \mathrm{H}_{2} \mathrm{O}$, Validyne, Northridge, California). Volume was determined via digital integration of the flow signal. A differential pressure transducer (MP-46 $\pm 100 \mathrm{~cm} \mathrm{H} \mathrm{H}_{2} \mathrm{O}$, Validyne, Northridge, California) was used to measure airway opening pressure. Cardiac output and pulmonary arterial pressure were measured via a 7.5 French pulmonary artery catheter (831 HF 7.5, Edwards Life Sciences, Irvine, California) inserted into the left external jugular vein. Proper position of the catheter was confirmed via pressure waveform analysis before and after balloon occlusion. Following instrumentation and a $30-\mathrm{min}$ stabilization period, 5 transport ventilators were randomly applied.

All signals (flow at the ETT, airway opening pressure, arterial blood pressure, and pulmonary arterial pressure) were amplified (8805C, Hewlett Packard, Waltham, Massachusetts), converted to digital signals with an analog-to-
digital converter (DI-220, Dataq Instruments, Akron, Ohio), and recorded at a sampling rate of 100 Hz , with dataacquisition software (Windaq/200, version 1.36, Dataq Instruments, Akron, Ohio). Ventilatory measurements made during the animal tests were all made at body-tem-perature-and-pressure-saturated conditions. All infusions, including the anesthetic, were given via volumetric infusion pump. A heating blanket was used to maintain a core temperature of $38-39^{\circ} \mathrm{C}$. An orogastric tube was placed to empty the stomach.

## Lung Injury

Severe lung injury was produced with bilateral lung lavage via instillations of 1 L of isotonic saline, warmed to $39^{\circ} \mathrm{C}$, repeated every 30 min , until $\mathrm{P}_{\mathrm{aO}_{2}}$ decreased to $\leq 100 \mathrm{~mm} \mathrm{Hg}$ at an $\mathrm{F}_{\mathrm{IO}_{2}}$ of 1.0 and a PEEP of $5 \mathrm{~cm} \mathrm{H}_{2} \mathrm{O}$. A stable lung injury was defined as a $\mathrm{P}_{\mathrm{aO}}$, change of $<10 \%$ after 60 min . It took 2-4 lavages and 2-3 hours to establish a stable lung injury. During development and stabilization of lung injury, the animals were ventilated with the Puritan Bennett 840 ventilator. After a stable lung injury was established, 5 transport ventilators were randomly applied.

On completion of the protocol, the animals were sacrificed under deep anesthesia ( $10 \mathrm{mg} / \mathrm{kg}$ pentobarbital) with rapid infusion of 50 mL saturated potassium chloride solution. Electrocardiogram and arterial blood pressure readings confirmed cardiac standstill.

## Statistical Analysis

Formal statistical analysis was not performed. Lung model data were compared to the ventilator settings. A difference $>10 \%$ was considered excessive, because most of the ventilator manufacturers indicate that the normal range of operation is within $10 \%$ of the set parameters. The mean $\pm \mathrm{SD} \mathrm{V}_{\mathrm{T}}$ was calculated from 5 breaths.

PIP, PEEP, and RR did not change with any ventilator during the bench evaluation. During the normal and in-jured-lung animal evaluations, the ability of each ventilator to achieve the target blood gas values was evaluated. The oxygen cylinder duration and battery life were recorded in minutes.

## Results

## Bench Test

The 15 ventilators evaluated can be classified as either "simple" or "sophisticated" transport ventilators, and as those that require compressed gas (pneumatic), those that can operate without compressed gas but require electrical power, and those that require both or either power source. The data in the tables and figures are organized with that

Table 2. Operational Features of Evaluated Ventilators: Battery, Gas Consumption, Alarms, and Ease of Use

| Ventilator | Battery Powered | Battery Life* | External Gas Required | Oxygen <br> Cylinder <br> Duration <br> $(\mathrm{min}) \dagger$ | Disconnect | HighPressure Alarm | Able to <br> Ventilate <br> Normal <br> Lungs | Able to Ventilate Injured Lungs | $\begin{aligned} & \text { Ease } \\ & \text { of } \\ & \text { Use } \end{aligned}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Sophisticated |  |  |  |  |  |  |  |  |  |
| Univent Eagle 754 | Yes | 4 h | No | 35 | Yes | Yes | Yes | No | 1 |
| VersaMed iVent | Yes | 90 min | No | 52 | Yes | Yes | Yes | Yes | 1 |
| Newport HT50 | Yes | $8 \mathrm{~h}, 10 \mathrm{~min}$ | No | 46 | Yes | Yes | Yes | Yes | 1 |
| Pulmonetic Systems LTV 1000 | Yes | 75 min | No | 32 | Yes | Yes | Yes | No | 1 |
| Simple |  |  |  |  |  |  |  |  |  |
| Oceanic Medical Products Magellan | No | NA | Yes | 60 | Yes | Yes | Yes | No | 1 |
| Bio-Med Devices IC2A | No | NA | Yes | 30 | No | No | Yes | Yes | 2 |
| Pneupac Parapac Medic | No | NA | Yes | 688 | Yes | Yes | Yes | No | 1 |
| Pneupac Parapac Transport 200D | No | NA | Yes | 62 | Yes | Yes | Yes | No | 1 |
| Life Support Products AutoVent 2000 | No | NA | Yes | 60 | No | No | Yes | No | 1 |
| Carevent ATV + | No | NA | Yes | 65 | Yes | Yes | Yes | No | 1 |
| Vortran RespirTech Pro\\| | No | NA | Yes | Variable | No | No | No | No | 3 |
| Percussionaire TXP\|| | No | NA | Yes | 779 | No | No | Yes | Yes | 3 |
| Bio-Med Devices Crossvent 3** | Yes | NA | Yes | 53 | No | No | Yes | No | 1 |
| Bird Avian** | Yes | NA | Yes | 30 | Yes | Yes | Yes | No | 1 |
| Pneupac Compac 200 | Yes | 4 h | No | 65 | Yes | Yes | Yes | No | 2 |

[^3]schema. Five of the ventilators tested can be classified as sophisticated transport ventilators; the other ten are simple transport ventilators.

Eight of the ventilators were purely pneumatic (they rely completely on compressed gas and do not require any other power source). Most of these incorporate an airentrainment device to provide different $\mathrm{F}_{\mathrm{IO}_{2}}$ values. The Oceanic Medical Products Magellan and the Life Support Products AutoVent 2000 are the exceptions; all breaths are delivered with $100 \%$ source gas. Four ventilators were capable of ventilating with only battery or alternatingcurrent power. Oxygen was not required, but may be added to increase $\mathrm{F}_{\mathrm{IO}_{2}}$.

The third group consisted of the 3 ventilators that require both compressed gas and electricity (battery or alternating current). These ventilators incorporate a pneumatic gas-delivery system and also require battery or alternating current to operate their electronic controls.

Table 1 shows the ventilators' physical dimensions and ventilation modes. There are considerable differences in the size and weight. In some ventilators the modes are very limited, whereas others have multiple pressure and volume modes.

Gas consumption (Table 2) ranged from 30 min to 77 min .

Battery life ranged from 75 min to 8 hours and 10 min . All except 5 ventilators (Bio-Med Devices IC2A, Bio-Med Devices Crossvent 3, Life Support Products AutoVent 2000, Percussionaire TXP, and Vortran RespirTech Pro) incorporated both a low-pressure/disconnect alarm and a high-inspiratory-pressure alarm.

One interesting finding was with the Bio-Med Devices Crossvent 3. When changing from the "Air Mix" setting to $100 \%$ oxygen, the delivered $\mathrm{V}_{\mathrm{T}}$ approximately doubled above the set $\mathrm{V}_{\mathrm{T}}$. According to the manufacturer, this is expected, and the operations manual instructs to adjust the flow accordingly.

The Percussionaire TXP was the most difficult to operate, and its controls were not clearly identified. The BioMed Devices IC2 and the Oceanic Medical Products Magellan were the only ventilators operable near a magnetic resonance imaging device.

Tables 3 and 4 show the bench performance data. Most of the ventilators performed at or close to specifications under bench test conditions. The measured $\mathrm{V}_{\mathrm{T}}$ of most of the ventilators was less than the set $\mathrm{V}_{\mathrm{T}}$, and this discrepancy increased under conditions of increased resistance or decreased compliance. Seventy-eight individual tests were performed at a $\mathrm{V}_{\mathrm{T}}$ of $1,000 \mathrm{~mL}$. In 32 of these tests the
Table 3. Bench Performance at a Tidal Volume of 500 mL

|  | High Resistance and Normal Compliance |  |  |  | Normal Resistance and Normal Compliance |  |  |  | Normal Resistance and Low Compliance |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\begin{gathered} \mathrm{V}_{\mathrm{T}} \\ (\text { mean } \pm \\ \text { SD mL) } \\ \hline \end{gathered}$ | $\begin{gathered} \mathrm{RR} \\ (\mathrm{br} / \mathrm{min}) \end{gathered}$ | $\begin{gathered} \text { PIP } \\ \left(\mathrm{cm} \mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ | $\begin{gathered} \text { PEEP } \\ \left(\mathrm{cm} \mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ | $\begin{gathered} \mathrm{V}_{\mathrm{T}} \\ (\text { mean } \pm \\ \text { SD mL) } \\ \hline \end{gathered}$ | RR $(\mathrm{br} / \mathrm{min})$ | $\begin{gathered} \mathrm{PIP} \\ \left(\mathrm{~cm} \mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ | $\begin{gathered} \text { PEEP } \\ \left(\mathrm{cm} \mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ | $\begin{gathered} \mathrm{V}_{\mathrm{T}} \\ (\text { mean } \pm \\ \text { SD mL) } \end{gathered}$ | $\begin{gathered} \mathrm{RR} \\ (\mathrm{br} / \mathrm{min}) \end{gathered}$ | $\begin{gathered} \mathrm{PIP} \\ \left(\mathrm{~cm} \mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ | $\begin{gathered} \text { PEEP } \\ \left(\mathrm{cm} \mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ |
| Sophisticated Transport Ventilators |  |  |  |  |  |  |  |  |  |  |  |  |
| Univent Eagle 754* | $554 \pm 3.1 \dagger$ | 15 | 19 | 5 | $583 \pm 4.5 \dagger$ | 15 | 12 | 5 | $571 \pm 8.4 \dagger$ | 15 | 32 | 5 |
| Univent Eagle 754\% | $535 \pm 2.9$ | 29 | 31 | 5 | $570 \pm 5.1 \dagger$ | 30 | 16 | 5 | $555 \pm 3.6 \dagger$ | 29 | 34 | 5 |
| VersaMed iVent* | $557 \pm 3.2 \dagger$ | 15 | 19 | 4 | $541 \pm 11.5$ | 15 | 12 | 5 | $515 \pm 16.3$ | 15 | 32 | 4 |
| VersaMed iVent $\ddagger$ | $401 \pm 7.1 \dagger$ | 30 | 25 | 4 | $555 \pm 25.2 \dagger$ | 30 | 16 | 4 | $496 \pm 6.1$ | 30 | 32 | 5 |
| Newport HT50* | $480 \pm 9.2$ | 15 | 18 | 4 | $477 \pm 12.1$ | 15 | 14 | 4 | $498 \pm 4.5$ | 15 | 29 | 5 |
| Newport HT50\% | $468 \pm 4.8$ | 29 | 21 | 9 | $459 \pm 7.4$ | 30 | 14 | 5 | $494 \pm 6.7$ | 30 | 30 | 4 |
| Pulmonetic Systems <br> LTV 1000* | $483 \pm 3.5$ | 15 | 16 | 5 | $486 \pm 2.0$ | 15 | 14 | 5 | $476 \pm 3.5$ | 15 | 29 | 5 |
| Pulmonetic Systems LTV 1000 $\ddagger$ | $483 \pm 4.2$ | 30 | 20 | 7 | $484 \pm 3.8$ | 30 | 16 | 6 | $477 \pm 1.3$ | 30 | 29 | 5 |
| Simple Transport Ventilators |  |  |  |  |  |  |  |  |  |  |  |  |
| Oceanic Medical Products Magellan* | $411 \pm 1.3 \dagger$ | $23 \dagger$ | 16 | 5 | $462 \pm 2.9$ | $21 \dagger$ | 11 | 5 | $441 \pm 10.4 \dagger$ | $21 \dagger$ | 26 | 4 |
| Oceanic Medical Products Magellan $\ddagger$ | $428 \pm 3.1 \dagger$ | 30 | 26 | 5 | $410 \pm 0.9 \dagger$ | 31 | 12 | 5 | $418 \pm 1.6 \dagger$ | 32 | 27 | 5 |
| Bio-Med Devices IC2A* | $487 \pm 0.6$ | 17† | 17 | 6 | $487 \pm 0.8$ | 17† | 13 | 6 | $474 \pm 1.5$ | 18† | 31 | 6 |
| Bio-Med Devices $\mathrm{IC} 2 \mathrm{~A} \ddagger$ | $483 \pm 1.7$ | 30 | 31 | 7 | $519 \pm 1.8$ | 30 | 16 | 6 | $487 \pm 3.1$ | 30 | 32 | 6 |
| Pneupac Parapac Medic* | $507 \pm 1.0$ | 15 | 17 | 5 | $530 \pm 1.2$ | 15 | 16 | 6 | $483 \pm 2.9$ | 16 | 28 | 5 |
| Pneupac Parapac Medic $\ddagger$ | $306 \pm 0.9 \dagger$ | 30 | 14 | 6 | $324 \pm 1.0 \dagger$ | 30 | 13 | 6 | $312 \pm 2.6 \dagger$ | 30 | 20 | 5 |
| Pneupac Parapac <br> Transport 200D* | $472 \pm 1.6$ | 16 | 18 | 5 | $485 \pm 1.8$ | 16 | 14 | 5 | $462 \pm 10.2$ | 16 | 27 | 4 |
| Pneupac Parapac Transport 200D $\ddagger$ | $456 \pm 1.5$ | 30 | 20 | 7 | $479 \pm 0.8$ | 30 | 15 | 6 | $461 \pm 11.7$ | 30 | 27 | 5 |
| Life Support Products AutoVent 2000* | $475 \pm 1.1$ | 15 | 13 | NA | $504 \pm 2.7$ | 15 | 12 | NA | $445 \pm 7.5 \dagger$ | 15 | 23 | NA |
| Life Support Products AutoVent 2000 $\ddagger$ | $472 \pm 5.5$ | $17 \dagger$ | 12 | NA | $497 \pm 5.3$ | $18 \dagger$ | 12 | NA | $438 \pm 9.8 \dagger$ | $18 \dagger$ | 24 | NA |
| Carevent ATV+* | $535 \pm 1.3$ | $13 \dagger$ | 16 | 5 | $560 \pm 3.1 \dagger$ | $12 \dagger$ | 15 | 5 | $549 \pm 2.0$ | $12 \dagger$ | 31 | 6 |
| Carevent ATV $+\ddagger$ | $555 \pm 5.1 \dagger$ | 25 $\dagger$ | 24 | 5 | $570 \pm 3.8 \dagger$ | 25† | 16 | 5 | $575 \pm 10.7 \dagger$ | 25 $\dagger$ | 34 | 6 |
| Vortran RespirTech Pro | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA |
| Percussionaire TXP | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA |

Table 3. (Continued)

|  | High Resistance and Normal Compliance |  |  |  | Normal Resistance and Normal Compliance |  |  |  | Normal Resistance and Low Compliance |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\begin{gathered} \mathrm{V}_{\mathrm{T}} \\ (\text { mean } \pm \\ \text { SD mL) } \end{gathered}$ | $\begin{gathered} \mathrm{RR} \\ (\mathrm{br} / \mathrm{min}) \end{gathered}$ | $\begin{gathered} \mathrm{PIP} \\ \left(\mathrm{~cm} \mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ | $\left.\begin{array}{l} \text { PEEP } \\ (\mathrm{cm} \mathrm{H} \\ 2 \end{array}\right)$ | $\begin{gathered} \mathrm{V}_{\mathrm{T}} \\ (\text { mean } \pm \\ \text { SD mL) } \end{gathered}$ | $\begin{gathered} \mathrm{RR} \\ (\mathrm{br} / \mathrm{min}) \end{gathered}$ | $\begin{gathered} \text { PIP } \\ \left(\mathrm{cm} \mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ | $\begin{gathered} \text { PEEP } \\ \left(\mathrm{cm} \mathrm{H} \mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ | $\begin{gathered} \mathrm{V}_{\mathrm{T}} \\ (\mathrm{mean} \pm \\ \mathrm{SD} \mathrm{~mL}) \end{gathered}$ | $\begin{gathered} \mathrm{RR} \\ (\mathrm{br} / \mathrm{min}) \end{gathered}$ | $\begin{gathered} \mathrm{PIP} \\ \left(\mathrm{~cm} \mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ | $\begin{gathered} \text { PEEP } \\ \left(\mathrm{cm} \mathrm{H} \mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ |
| Bio-Med Devices Crossvent 3* | $526 \pm 2.0$ | 15 | 20 | 5 | $532 \pm 0.9$ | 15 | 16 | 5 | $507 \pm 1.1$ | 15 | 30 | 5 |
| Bio-Med Devices Crossvent $3 \ddagger$ | $507 \pm 1.5$ | 30 | 25 | 8 | $523 \pm 0.8$ | 30 | 18 | 9 | $520 \pm 1.1$ | 30 | 31 | 5 |
| Bird Avian* | $472 \pm 0.6$ | 15 | 15 | 6 | $467 \pm 3.2$ | 15 | 13 | 7 | $475 \pm 1.3$ | 15 | 29 | 6 |
| Bird Avian ${ }_{\text {\% }}$ | $473 \pm 1.6$ | 30 | 29 | 7 | $489 \pm 2.7$ | 30 | 16 | 10 | $488 \pm 2.9$ | 30 | 32 | 6 |
| Pneupac Compac 200* | $410 \pm 4.0 \dagger$ | 15 | 14 | 6 | $429 \pm 5.8 \dagger$ | 14 | 13 | 5 | $390 \pm 8.7 \dagger$ | 14 | 23 | 4 |
| Pneupac Compac 200 $\ddagger$ | $436 \pm 5.0 \dagger$ | 29 | 20 | 6 | $442 \pm 5.8 \dagger$ | 29 | 15 | 6 | $424 \pm 6.9 \dagger$ | 29 | 26 | 5 |

*Respiratory rate set at 15 breaths $/ \mathrm{min}$
$\dagger$ Measured value was $>10 \%$ different from the setting on the ventilator
$\ddagger$ Respiratory rate set at 30 breaths $/ \mathrm{min}$
$\stackrel{\text { Respiratory rate set at }}{\ddagger} \mathrm{V}_{\mathrm{T}}=$ tidal volume. $\mathrm{RR}=$ respiratory rate. $\mathrm{br} / \mathrm{min}=$ breaths $/ \mathrm{min} . \mathrm{PIP}=$ peak inspiratory pressure. $\mathrm{PEEP}=$ positive end-expiratory pressure. $\mathrm{NA}=$ not applicable.
measured $\mathrm{V}_{\mathrm{T}}$ was $>10 \%$ different than the set $\mathrm{V}_{\mathrm{T}}$, and in 19 of these tests the measured RR was $>10 \%$ different than the set RR.

Seventy-eight individual tests were performed at a $\mathrm{V}_{\mathrm{T}}$ of 500 mL . In 28 of these tests the measured $\mathrm{V}_{\mathrm{T}}$ was $>10 \%$ different than set $\mathrm{V}_{\mathrm{T}}$, and in 15 of these tests the RR was $>10 \%$ different than the set RR. The inability to meet the target $\mathrm{V}_{\mathrm{T}}$ is partially explained by compressible volume loss, since none of these units compensates for the volume loss due to compression. The compressible volume was $0.91 \pm 0.39 \mathrm{~mL} / \mathrm{cm} \mathrm{H}_{2} \mathrm{O}$ (range $0.43-1.65 \mathrm{~mL} / \mathrm{cm}$ $\mathrm{H}_{2} \mathrm{O}$ ), and since the PIP values were at most in the mid30s, only about one third of the volume loss can be attributed to compressible volume. Two of the ventilators (Vortran RespirTech Pro and Percussionaire TXP) were not included in the bench test because their design was incompatible with our protocol; they did not incorporate readily identifiable parameter settings.

Three of the ventilators were unable to meet some of the test conditions, due to their designs. The Carevent ATV + achieves $V_{T}$ by setting $\dot{\mathrm{V}}_{\mathrm{E}}$ and $R R$, with a maximum $\dot{\mathrm{V}}_{\mathrm{E}}$ of $14 \mathrm{~L} / \mathrm{min}$ and an inspiratory-expiratory ratio of $1: 2$ to establish $\mathrm{T}_{\mathrm{I}}$. The Pneupac Compac 200 also achieves $\mathrm{V}_{\mathrm{T}}$ by setting $\dot{\mathrm{V}}_{\mathrm{E}}$ and RR with a maximum $\dot{\mathrm{V}}_{\mathrm{E}}$ of $14 \mathrm{~L} / \mathrm{min}$. The Life Support Products AutoVent 2000 has 2 controls: $\mathrm{V}_{\mathrm{T}}$ and RR, with a maximum RR of $17-18$ breaths/min. Inspiratory time and flow vary with changes in $\dot{\mathrm{V}}_{\mathrm{E}}$. At high $R R$ settings the inspiratory-expiratory ratio is $>1: 1$.

## Animal Evaluations

We evaluated 14 ventilators with healthy and saline-lavage-injured sheep. We were unable to evaluate the Vortran RespirTech Pro in either animal model because the weight of the sheep ( $20-31 \mathrm{~kg}$ ) was below the operating range of this ventilator (minimum 40 kg ). All of the 14 ventilators tested were capable of ventilating the healthy lungs (Fig. 1). Ventilators without $\mathrm{F}_{\mathrm{IO}_{2}}$ control met the pH and $\mathrm{P}_{\mathrm{aCO}_{2}}$ targets, but exceeded the $\mathrm{P}_{\mathrm{aO}_{2}}$ target. With all the ventilators, hemodynamics were stable throughout the tests.

Four ventilators (VersaMed iVent, Newport HT50, BioMed Devices IC2A, and Percussionaire TXP) met all the targets when ventilating injured lungs (Fig. 2). Seven ventilators (Pulmonetic Systems LTV 1000, Bird Avian, Oceanic Medical Products Magellan, Pneupac Parapac Transport 200D, Pneupac Parapac Medic, Bio-Med Devices Crossvent 3, and Carevent ATV+) successfully ventilated the lungs of only one of the 2 lung-injured animals. The remaining 3 ventilators (Univent Eagle 754, Pneupac Compac 200, and Life Support Products AutoVent 2000) failed to meet the ventilation targets in either of the 2 lunginjured sheep used. In all cases of failure, the $\mathrm{P}_{\mathrm{aCO}_{2}}$ and/or pH targets were not met. In most cases, the RR setting was the limiting parameter. All 14 ventilators met the oxygen-
Table 4. Bench Performance at a Tidal Volume of $1,000 \mathrm{~mL}$

|  | High Resistance and Normal Compliance |  |  |  | Normal Resistance and Normal Compliance |  |  |  | Normal Resistance and Low Compliance |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\begin{gathered} \stackrel{V_{\mathrm{T}} \mathrm{M}}{\left(\mathrm{mean}^{ \pm}\right.} \\ \mathrm{SD} \mathrm{~mL}) \end{gathered}$ | $\begin{aligned} & \mathrm{RR} \\ & (\mathrm{br} / \\ & \mathrm{min}) \end{aligned}$ | $\begin{gathered} \text { PIP } \\ (\mathrm{cm} \\ \left.\mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ | $\begin{aligned} & \text { PEEP } \\ & (\mathrm{cm} \\ & \left.\mathrm{H}_{2} \mathrm{O}\right) \end{aligned}$ |  | $\begin{aligned} & \mathrm{RR} \\ & (\mathrm{br} / \\ & \mathrm{min}) \end{aligned}$ | $\begin{gathered} \mathrm{PIP} \\ (\mathrm{~cm} \\ \left.\mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ | PEEP (cm $\mathrm{H}_{2} \mathrm{O}$ ) |  | $\begin{gathered} \mathrm{RR} \\ (\mathrm{br} / \\ \mathrm{min}) \end{gathered}$ | $\begin{gathered} \mathrm{PIP} \\ (\mathrm{~cm} \\ \left.\mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ | $\begin{gathered} \text { PEEP } \\ (\mathrm{cm} \\ \left.\mathrm{H}_{2} \mathrm{O}\right) \end{gathered}$ |
| Sophisticated Transport Ventilators |  |  |  |  |  |  |  |  |  |  |  |  |
| Univent Eagle 754* | $1,028 \pm 4.7$ | 10 | 36 | 5 | $1,102 \pm 3.1 \dagger$ | 10 | 22 | 5 | $1,055 \pm 4.7$ | 10 | 56 | 5 |
| Univent Eagle 754¥ | $1,027 \pm 2.8$ | 20 | 35 | 5 | $1,097 \pm 6.8$ | 20 | 22 | 5 | $1,009 \pm 5.7$ | 20 | 55 | 5 |
| VersaMed iVent* | $989 \pm 74.9$ | 10 | 33 | 5 | $996 \pm 58.8$ | 10 | 20 | 5 | $906 \pm 17.3$ | 10 | 51 | 5 |
| VersaMed iVent $\ddagger$ | $932 \pm 6.3$ | 20 | 40 | 5 | $972 \pm 66.6$ | 20 | 20 | 4 | $929 \pm 0.8$ | 20 | 51 | 5 |
| Newport HT50* | $926 \pm 8.9$ | 10 | 36 | 4 | $940 \pm 24.9$ | 10 | 23 | 3 | $962 \pm 20.2$ | 10 | 52 | 5 |
| Newport HT50\% | $920 \pm 14.8$ | 20 | 37 | 5 | $944 \pm 14.4$ | 20 | 24 | 4 | $967 \pm 15.4$ | 20 | 54 | 5 |
| Pulmonetic Systems LTV 1000* | $955 \pm 1.1$ | 10 | 36 | 5 | $945 \pm 6.7$ | 10 | 24 | 5 | $927 \pm 10.8$ | 10 | 52 | 5 |
| Pulmonetic Systems LTV 1000\% | $954 \pm 2.5$ | 20 | 37 | 6 | $949 \pm 1.7$ | 20 | 24 | 5 | $926 \pm 2.1$ | 20 | 52 | 5 |
| Simple Transport Ventilators |  |  |  |  |  |  |  |  |  |  |  |  |
| Oceanic Medical Products Magellan* | $789 \pm 2.2 \dagger$ | $12 \dagger$ | 29 | 5 | $752 \pm 1.6 \dagger$ | $12 \dagger$ | 16 | 4 | $753 \pm 6.1 \dagger$ | $12 \dagger$ | 42 | 4 |
| Oceanic Medical Products Magellan | $870 \pm 3.3 \dagger$ | $24 \dagger$ | 34 | 6 | $842 \pm 4.2 \dagger$ | $25 \dagger$ | 17 | 5 | $850 \pm 7.6 \dagger$ | $25 \dagger$ | 47 | 5 |
| Bio-Med Devices IC2A* | $910 \pm 1.7$ | 9 | 35 | 6 | $952 \pm 2.1$ | 10 | 20 | 6 | $898 \pm 4.4 \dagger$ | 10 | 52 | 6 |
| Bio-Med Devices IC2A $\ddagger$ | $887 \pm 1.2 \dagger$ | 21 | 37 | 7 | $937 \pm 1.1$ | 21 | 20 | 6 | $886 \pm 4.1 \dagger$ | 21 | 52 | 6 |
| Pneupac Parapac Medic* | $942 \pm 2.9$ | 11 | 28 | 5 | $973 \pm 3.5$ | 11 | 24 | 6 | $899 \pm 6.5 \dagger$ | 10 | 53 | 5 |
| Pneupac Parapac Medic $\ddagger$ | $635 \pm 2.0$ | 20 | 24 | 5 | $653 \pm 2.2 \dagger$ | 20 | 19 | 6 | $614 \pm 4.6 \dagger$ | 20 | 36 | 5 |
| Pneupac Parapac Transport 200D* | $784 \pm 2.7 \dagger$ | $12 \dagger$ | 27 | 5 | $840 \pm 1.7 \dagger$ | $12 \dagger$ | 21 | 4 | $750 \pm 15.5 \dagger$ | $12 \dagger$ | 41 | 4 |
| Pneupac Parapac Transport 200D $\ddagger$ | $774 \pm 1.6 \dagger$ | 21 | 28 | 6 | $835 \pm 2.4 \dagger$ | 21 | 21 | 5 | $755 \pm 3.2 \dagger$ | $22 \dagger$ | 42 | 5 |
| Life Support Products AutoVent 2000* | $925 \pm 1.5$ | 10 | 23 | NA | $948 \pm 4.7$ | 10 | 20 | NA | $892 \pm 2.6 \dagger$ | 10 | 44 | NA |
| Life Support Products AutoVent 2000 $\ddagger$ | $913 \pm 4.8$ | $17 \dagger$ | 22 | NA | $939 \pm 5.4$ | $17 \dagger$ | 20 | NA | $896 \pm 4.1 \dagger$ | $17 \dagger$ | 44 | NA |
| Carevent ATV+* | $1,132 \pm 1.9 \dagger$ | 9 | 28 | 5 | $1,164 \pm 1.3 \dagger$ | 9 | 25 | 5 | $1,125 \pm 5.2 \dagger$ | 9 | 56 | 6 |
| Carevent ATV+ұ | $1,142 \pm 1.3 \dagger$ | $13 \dagger$ | 33 | 5 | $1,179 \pm 5.2 \dagger$ | $13 \dagger$ | 26 | 5 | $1,149 \pm 1.4 \dagger$ | $13 \dagger$ | 58 | 5 |
| Vortran RespirTech Pro | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA |
| Percussionaire TXP | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA |
| Bio-Med Devices Crossvent 3* | $975 \pm 0.7$ | 10 | 38 | 4 | $1,016 \pm 1.7$ | 10 | 25 | 5 | $948 \pm 1.1$ | 10 | 51 | 5 |
| Bio-Med Devices Crossvent $3 \ddagger$ | $972 \pm 0.9$ | 20 | 39 | 5 | $1,017 \pm 0.8$ | 20 | 25 | 4 | $953 \pm 9.7$ | 20 | 51 | 4 |
| Bird Avian* | $950 \pm 2.6$ | 10 | 33 | 6 | $970 \pm 3.1$ | 10 | 20 | 6 | $958 \pm 4.7$ | 10 | 52 | 5 |
| Bird Avian $\ddagger$ | $947 \pm 2.7$ | 20 | 35 | 8 | $972 \pm 3.1$ | 20 | 21 | 7 | $958 \pm 4.0$ | 20 | 53 | 6 |
| Pneupac Compac 200* | $917 \pm 1.3$ | 9 | 24 | 6 | $917 \pm 9.8$ | 9 | 21 | 5 | $794 \pm 13.6 \dagger$ | 9 | 42 | 4 |
| Pneupac Compac 200\% | $654 \pm 6.0 \dagger$ | $18 \dagger$ | 23 | 6 | $654 \pm 7.3 \dagger$ | $18 \dagger$ | 15 | 5 | $629 \pm 12.2 \dagger$ | $18 \dagger$ | 36 | 4 |

*Respiratory rate set at 15 breaths/min
$\dagger$ Measured value was $>10 \%$ different fi
*Respiratory rate set at 15 breaths/min
$\dagger$ Measured value was $>10 \%$ different from the setting on the ventilator
$\ddagger$ Respiratory rate set at 30 breaths $/ \mathrm{min}$
$\mathrm{V}_{\mathrm{T}}=$ tidal volume. $\mathrm{RR}=$ respiratory rate. $\mathrm{br} / \mathrm{min}=$ breaths $/ \mathrm{min} . \mathrm{PIP}=$ peak inspiratory pressure. $\mathrm{PEEP}=$ positive end-expiratory pressure. $\mathrm{NA}=$ not applicable.


Fig. 1. $\mathrm{P}_{\mathrm{acO}}^{2}$, arterial pH , and ratio of $\mathrm{P}_{\mathrm{aO}_{2}}$ to fraction of inspired oxygen ( $\mathrm{F}_{\mathrm{IO}_{2}}$ ) in healthy (no lung injury) sheep during ventilation with 14 transport ventilator models. Each set of values represents data from a single sheep. Assessment was performed on 2 sheep with each ventilator. The large variability in the ratio of $\mathrm{P}_{\mathrm{aO}_{2}}$ to fraction of inspired oxygen $\left(\mathrm{P}_{\mathrm{aO}_{2}} / \mathrm{F}_{\mathrm{IO}_{2}}\right)$ is because some of the ventilators only offer only 1 or $2 \mathrm{~F}_{1 \mathrm{O}_{2}}$ settings, and because of the level of gas exchange in each sheep. The Vortran RespirTech Pro could not be used on the animals we tested. The Percussionaire TXP and the Vortran RespirTech Pro could not be set to the specifications required by the lung model. $\mathrm{U}=$ Univent Eagle 754. P = Pulmonetic Systems LTV 1000. V = VersaMed iVent. B = Bird Avian. M = Oceanic Medical Products Magellan. $\mathrm{N}=$ Newport HT50. PT = Pneupac Parapac Transport 200D. PM = Pneupac Parapac Medic. $\mathrm{C}=$ Pneupac Compac 200. $\mathrm{BI}=$ Bio-Med Devices IC2A. BC = Bio-Med Devices Crossvent 3. CV = Carevent ATV + . A = Life Support Products AutoVent 2000. PC = Percussionaire TXP.
ation target under all conditions with the lung-injured sheep. As with the uninjured sheep, with all the ventilators the hemodynamics were stable throughout the tests.

## Discussion

The major findings of this study are:

1. All the evaluated ventilators were able to maintain normal ventilation and hemodynamics in healthy sheep.
2. In the lung-injured sheep, few of the ventilators could be set to meet the $\mathrm{P}_{\mathrm{aCO}_{2}}$ or pH targets. The ventilators unable to meet these targets were limited by the RR setting.
3. In the bench study, only 6 of the ventilators met the $\mathrm{V}_{\mathrm{T}}$ and RR settings under all the test conditions.
4. Only 5 of the ventilators (Univent Eagle 754, VersaMed iVent, Newport HT50, Pulmonetic Systems LTV 1000, and Pneupac Compac 200) can operate without a compressed gas source, and their battery life differed considerably.
5. A full E-size cylinder of oxygen allowed ventilation with $100 \%$ oxygen for only $30-77 \mathrm{~min}$.
6. The 2 ventilators most suitable for use in front-line rescue situations, where oxygen may not be available, are the Newport HT50 and the Univent Eagle 754.

## Use of Transport Ventilators

Transport ventilators are required in various settings: intra-hospital, inter-hospital, pre-hospital, and in the field by military or civilian authorities. ${ }^{3}$ Each of these settings has different priorities regarding ventilator design. In forward military or field use by civilian groups, the ideal ventilator would be simple to operate, battery powered, compact, lightweight, and would operate without compressed gas. In that setting it is unlikely that the patient will be breathing spontaneously, so versatility of available modes is unnecessary. Similar issues exist during pre-hospital transport, but compressed gas is readily available in most ambulances, so a pneumatically operated ventilator is as acceptable as a battery operated unit. During inter-hospital transport the patient may be breathing spontaneously, which necessitates patient-triggered ventilation, and frequently these patients require high $\mathrm{F}_{\mathrm{IO}_{2}}$.

The most common use of transport ventilators is in in-tra-hospital transport. At Massachusetts General Hospital, the respiratory care department performs about 30 oneway patient transports per day and another $10-15$ are performed by the anesthesia department, all of which require continuous mechanical ventilation. Most of these trans-


Fig. 2. $\mathrm{P}_{\mathrm{aCO}_{2}}$, arterial pH , and ratio of $\mathrm{P}_{\mathrm{aO}_{2}}$ to fraction of inspired oxygen $\left(\mathrm{F}_{\mathrm{IO}_{2}}\right)$ in lung-injured sheep during ventilation with 14 transport ventilator models. Each set of values represents data from a single sheep. Assessment was performed on 2 sheep with each ventilator. The large variability in the ratio of $\mathrm{P}_{\mathrm{aO}}$ to fraction of inspired oxygen $\left(\mathrm{P}_{\mathrm{aO}_{2}} / \mathrm{F}_{\mathrm{IO}_{2}}\right)$ is because some of the ventilators only offer 1 or $2 \mathrm{~F}_{\mathrm{IO}_{2}}$ settings, and because of the level of gas exchange in each sheep. The Vortran RespirTech Pro could not be used on the animals we tested. The Percussionaire TXP and the Vortran RespirTech Pro could not be set to the specifications required by the lung model. U = Univent Eagle 754. $\mathrm{P}=$ Pulmonetic Systems LTV 1000. $\mathrm{V}=$ VersaMed iVent. $\mathrm{B}=$ Bird Avian. $\mathrm{M}=$ Oceanic Medical Products Magellan. $\mathrm{N}=$ Newport HT50. PT = Pneupac Parapac Transport 200D. PM = Pneupac Parapac Medic. C = Pneupac Compac 200. BI = Bio-Med Devices IC2A. BC = Bio-Med Devices Crossvent 3. CV = Carevent ATV+. A = Life Support Products AutoVent 2000. PC = Percussionaire TXP.
ports are to and from diagnostic areas, the operating room, or the emergency department.

It is well documented that transport ventilators provide more stable gas exchange and hemodynamics than manual ventilators. ${ }^{10-13}$ Gervais et al ${ }^{11}$ observed severe respiratory alkalosis during transport with manual ventilation. In 20 patients transported to diagnostic areas, Braman et al ${ }^{12}$ found substantial respiratory acidosis or alkalosis and hemodynamic compromise in 16 patients receiving manual ventilation. Hurst et al $^{13}$ also documented respiratory alkalosis during intrahospital transport of 28 patients receiving manual ventilation. Nakamura et al ${ }^{10}$ also observed greater variability of gas exchange and hemodynamics during transport with manual ventilation than with a transport ventilator.

## Types of Transport Ventilators

Austin et $\mathrm{al}^{3}$ classified transport ventilators into 3 categories, based on their capabilities: automatic resuscita-
tors, simple transport ventilators, and sophisticated transport ventilators. They defined a simple transport ventilator as one that provides a specified rate and volume with a high-pressure relief valve, ${ }^{3}$ whereas a sophisticated transport ventilator has modes that allow spontaneous breathing, and additional alarms and monitors of gas delivery. Five of the ventilators we evaluated were sophisticated: Newport HT50, Univent Eagle 754, VersaMed iVent, Pulmonetic Systems LTV 1000, Bird Avian.

We considered the Newport HT50 and the Univent Eagle 754 most suited for use in forward military positions, because they have longer battery life (8 hours and 4 hours, respectively). The VersaMed iVent was the heaviest of the units evaluated ( 10 kg ). However, the Newport HT50, Univent Eagle 754, VersaMed iVent, and Pulmonetic Systems LTV 1000 clearly could function exceptionally well in all transport settings if they had longer battery life. The Bird Avian was the most
limited in this regard, because it lacks a battery and needs compressed gas to operate.

## Issues/Problems With Specific Ventilators

The choice of a ventilator is also determined by other specific design issues. Only 2 of the ventilators (Bio-Med Devices IC2 and Oceanic Medical Products Magellan) are designed for use during magnetic resonance imaging. The following ventilators had no alarms: Vortran RespirTech Pro, Bio-Med Devices IC2, Percussionaire TXP, Oceanic Medical Products Magellan, and Life Support Products AutoVent 2000.
Many of the ventilators allow very few $\mathrm{F}_{\mathrm{IO}_{2}}$ values: Vortran RespirTech Pro ( $\mathrm{F}_{\mathrm{IO}_{2}} 1.0$ ), Bio-Med Devices IC2 ( $\mathrm{F}_{\mathrm{IO}_{2}}$ 1.0), Oceanic Medical Products Magellan $2000\left(\mathrm{~F}_{\mathrm{IO}_{2}} 1.0\right)$, Pneupac Parapac Transport 200D ( $\mathrm{F}_{\mathrm{IO}_{2}} 0.5$ or 1.0), Pneupac Parapac Medic ( $\mathrm{F}_{\mathrm{IO}_{2}} 0.5$ or 1.0 ), Bio-Med Devices Crossvent $3\left(\mathrm{~F}_{\mathrm{IO}_{2}} 0.5\right.$ or 1.0), Carevent ATV $+\left(\mathrm{F}_{\mathrm{IO}_{2}} 0.8\right.$ or 1.0), and Life Support Products AutoVent $2000\left(\mathrm{~F}_{\mathrm{IO}_{2}} 1.0\right)$.

The oxygen cylinder life of the Pneupac Parapac Medic exceeded the maximum estimated time ( 66 min ), because $\mathrm{V}_{\mathrm{T}}$ gradually decreased as the cylinder became depleted to 200 mL just before the ventilator shut down.

With the Percussionaire TXP, the maximum $\mathrm{F}_{\mathrm{IO}_{2}}$ delivered was 0.5 , which accounts for its $77-\mathrm{min}$ cylinder life. Note, however, that the volume of gas in E-size cylinders does vary, because filling pressure varies, which adds to the variability in cylinder life.

With the Newport HT50 and its nondisposable proprietary circuit, intrinsic PEEP developed at higher RR because of high expiratory resistance. With the Bio-Med Devices IC2, Life Support Products Magellan 2000, Pneupac Parapac Transport 200D, and Pneupac Parapac Medic the $V_{T}$ is set with the flow rate and $T_{I}$ controls, and $R R$ is controlled by those two plus an expiratory time control. With the Carevent ATV,$+ \mathrm{V}_{\mathrm{T}}$ is determined by $\dot{\mathrm{V}}_{\mathrm{E}}$ and RR. The Life Support Products AutoVent 2000 has 2 controls ( $R R$ and $V_{T}$ ), its maximum $R R$ is 18 breaths $/ \mathrm{min}$, and it does not have any alarms. The Pneupac Compac 200 is designed for military use. It has a sturdy case, and $V_{T}$ is adjusted by setting $\dot{V}_{E}$ and RR. It has a fixed $T_{I}$ of 1 s and a maximum RR of 26 breaths $/ \mathrm{min}$. The Percussionaire TXP is a pressure-limited and time-cycled ventilator, and its $\mathrm{V}_{\mathrm{T}}$ varied with changes in impedance, but we found that even with constant impedance the $\mathrm{V}_{\mathrm{T}}$ drifted upwards.
The maximum RR with the Life Support Products AutoVent 2000 is 18 breaths $/ \mathrm{min}$, and with the Carevent ATV+ it is 40 breaths $/ \mathrm{min}$. With the Oceanic Medical Products Magellan, setting RR at 15 breaths $/ \mathrm{min}$ and 20 breaths $/ \mathrm{min}$ resulted in measured RR of 23 breaths $/ \mathrm{min}$ and 30 breaths $/ \mathrm{min}$, respectively.

The most difficult ventilator to evaluate was the Vortran RespirTech Pro. This ventilator has few clearly labeled
controls, and the manufacturer's specified patient-weight range was outside the weight range of the animals we used. However, it is the smallest and lightest of the ventilators we tested, and it is only for single-patient use.

It may be necessary with some of these ventilators to monitor gas delivery with a secondary monitor because of the large difference between the set and actual $\mathrm{V}_{\mathrm{T}}$ and RR . Since we did not assess these ventilators during spontaneous breathing, we cannot comment on patient-ventilator synchrony or the difference between the set and delivered parameters during spontaneous ventilation.

## Comparison With Other Studies

Nolan et al ${ }^{5}$ evaluated the performance of 6 pneumatically operated ventilators. Similar to our results, they noted that the overall ability of the ventilators they tested to maintain delivered $\mathrm{V}_{\mathrm{T}}, \dot{\mathrm{V}}_{\mathrm{E}}$, and RR consistent with the set levels diminished as resistance increased or compliance decreased. McGough et al ${ }^{6}$ observed the same problem with 8 pneumatically operated ventilators they evaluated with a test lung. The Univent 750 was evaluated by Campbell et al, ${ }^{7}$ with a test lung, during controlled and patienttriggered ventilation. They observed, as we did, that with the Univent Eagle 754, gas delivery was not markedly affected by a decrease in compliance or an increase in resistance.

More recently, Miyoshi et al ${ }^{8}$ evaluated 4 ventilators with transport capabilities, all with internal batteries. However, at least 3 of these units (Puritan Bennett 740, Bird T-Bird, and Respironics Espirit) would not be considered typical transport ventilators. However, all of these units, along with the Pulmonetic Systems LTV 1000, were capable of ventilating a test lung during assisted ventilation, at various ventilation settings.

Zanetta et al ${ }^{9}$ evaluated 5 transport ventilators and 3 intensive care unit ventilators during controlled and pa-tient-triggered ventilation with a test lung. They determined that $\mathrm{V}_{\mathrm{T}}$ varied $<10 \%$ as delivered $\mathrm{V}_{\mathrm{T}}$ varied from 300 mL to 800 mL and compliance and resistance were varied. However, they noted that, because of high resistance to exhalation, all the portable ventilators they evaluated trapped gas at high $\dot{\mathrm{V}}_{\mathrm{E}}$.

## Limitations

The primary limitation of the present study is that it was not performed with patients. However, the bench and animal evaluations did simulate common settings required by patients during controlled ventilation. In addition, the animal model evaluations were consistent with pediatric patients, not adults. This limited the assessment of some of the ventilators. We also did not evaluate any of these ventilators during spontaneous breathing, which is clearly
a major issue in transport within and between hospitals. As a result, we cannot comment on their performance during spontaneous triggering. Also, we evaluated only one ventilator from each company, and we cannot be sure that the single ventilator we tested reflects the operation of all ventilators of that model.

## Conclusions

Only 2 of the transport ventilators evaluated met the trial targets in all bench and animal settings. With some ventilators the settings were inaccurate. The physical characteristics and high gas consumption of some of these ventilators may render them less desirable for patient transport.

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[^2]:    $\mathrm{F}_{\mathrm{IO}_{2}}=$ fraction of inspired oxygen
    PEEP $=$ positive end-expiratory pressure
    $\mathrm{A} / \mathrm{C}=$ assist/control
    SIMV $=$ synchronized intermittent mandatory ventilation
    $\mathrm{VP}=$ volume-controlled and pressure-controlled modes available
    PSV $=$ pressure support ventilation
    $\mathrm{CPAP}=$ continuous positive airway pressure
    CMV $=$ controlled mechanical ventilation
    IMV $=$ intermittent mandatory ventilation

[^3]:    *Battery life is based on tidal volume $\left(\mathrm{V}_{\mathrm{T}}\right)$ of 1 L , respiratory rate of 10 breath $/ \mathrm{min}$, and fraction of inspired oxygen $\left(\mathrm{F}_{\mathrm{IO}_{2}}\right)$ of 0.21 .
    $\dagger$ The oxygen cylinder duration is the time it took to consume 1 full E-size oxygen cylinder with the ventilator set at $\mathrm{V}_{\mathrm{T}}$ of 1 L , respiratory rate of 10 breaths/min, and $\mathrm{F}_{\mathrm{IO}}^{2}$ of 1.0
    $\ddagger$ Ease of use: $1=$ clearly labeled and easy to access; $2=$ clearly labeled but difficult to access; $3=$ not clearly labeled and difficult to access.
    $\S \mathrm{V}_{\mathrm{T}}$ gradually decreased as cylinder became depleted to 200 mL just before the ventilator shut down.
    $\|$ True pressure-cycled ventilator. Changes in resistance or compliance significantly altered the delivered $\mathrm{V}_{\mathrm{T}}$. Difficult to set at desired parameters.
    $T \mathrm{~F}_{\mathrm{IO}_{2}}$ fixed at 0.5 .
    **Pneumatically powered, electronically controlled ventilator. Battery and/or alternating current electricity are required for electronic controls, monitoring, etc. Compressed gas is required for ventilation.
    $\mathrm{NA}=$ not applicable

