

Reversed Halo Sign: A Systematic Review

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A reversed halo sign (RHS) is defined as the presence of a focal ring-shaped area of ground-glass opacity within a peripheral rim of consolidation. Although originally described in patients with cryptogenic organizing pneumonia, it has been described with several other noninfectious and infectious diseases, including fungal infections. Thus, it is imperative that a proper diagnosis be established before initiating treatment. In this article, we systematically review the literature (PubMed and Embase) for the associations of the RHS. We have also proposed a diagnostic algorithm for an approach to a patient with an RHS. Key words: reversed halo sign; RHS; computed tomography; HRCT; MDCT; *Aspergillus*; *mucormycosis*. [Respir Care 2014;59(9):1–•. © 2014 Daedalus Enterprises]

Introduction

The presence of central annular ground-glass opacity surrounded by a peripheral rim of consolidation, the so-called reversed halo sign (RHS), was first described in patients with cryptogenic organizing pneumonia¹ and was considered fairly specific for this disease.² Subsequently, several case reports and case series have reported the occurrence of the RHS in several other diseases (both infectious and noninfectious), thereby questioning the specificity of this sign. In this article, we systematically review the literature for the association of the RHS with various disorders. Furthermore, we also provide an algorithmic approach for a patient presenting with an RHS.

Methods

We first searched the PubMed and Embase databases for any systematic review reporting the association of the RHS with various diseases. Only narrative reviews were

found. We then independently searched the PubMed and Embase databases (January 1993 to November 2013) using the following free text terms: “reversed halo sign” OR “reverse halo sign” OR “atoll sign” OR “fairy ring sign” without any language restriction (Fig. 1). We included studies (case reports and original articles) describing the association of the RHS with various diseases. We excluded editorials and reviews. Any disagreement was resolved by discussion between the authors. The database created from the electronic searches was compiled in a reference manager package (EndNote X7, Thomson Reuters, Philadelphia, Pennsylvania), and all duplicate citations were eliminated. The citations were first screened by both authors to capture relevant studies. The full text of each citation was then obtained and reviewed in detail.

Data were recorded on a standard data extraction form. The following items were extracted: (1) publication details (title, authors, and other citation details), including country where the study was conducted; (2) patient demographics (age, sex, and immune status of the patient); and (3) modality used for establishing the diagnosis.

An institutional review board clearance was not required for this study, as this was a systematic review of published studies.

Results

Our initial database search yielded 94 citations, of which 58 case reports or case series (209 patients with an RHS) were included in the final review (Table 1).^{1–58} The RHS has been demonstrated to be present in several diseases, infectious as well as noninfectious (Table 2). The largest

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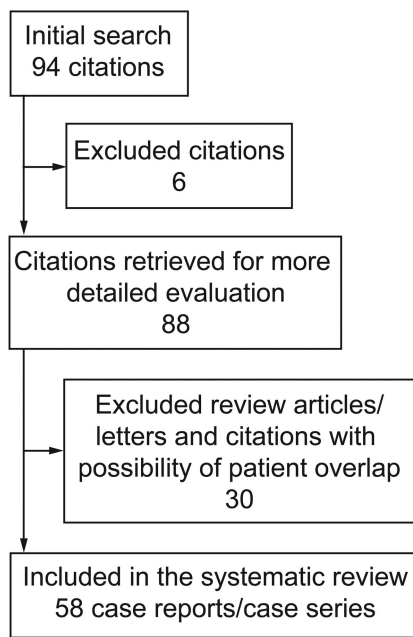


Fig. 1. Citation selection process for systematic review.

case series was published by Marchiori et al⁴⁶ and included 79 patients.

Infectious Diseases

The RHS is most commonly associated with invasive fungal infections, most commonly with mucormycosis (see Table 2). It has not been described in candidiasis or fusariosis. All the case reports of invasive fungal infections with an RHS are in immunocompromised hosts (hematologic malignancies, diabetes mellitus, and post-chemotherapy state). The RHS has also been described in endemic infections such as paracoccidioidomycosis and tuberculosis. In invasive fungal infection complicating immunosuppressed individuals, the sign appears as one or more large lesions, whereas in endemic mycoses, the lesions are bilateral and asymmetric, associated with ground-glass opacities, centrilobular nodules, or areas of consolidation.^{8,41} Other infections where it is uncommonly seen are histoplasmosis, cryptococcosis, influenza, psittacosis, *Pneumocystis*, and *Legionella* pneumonia. In patients with pneumococcal pneumonia, the RHS has been seen in the resolution phase and thus represents a post-infectious organizing pneumonia.³³

Noninfectious Diseases

The RHS was first described in cryptogenic organizing pneumonia, and this sign is most commonly associated with this condition. The RHS has been described in organizing pneumonia secondary to various causes (see Table

1). Other noninfectious diseases associated with the RHS are sarcoidosis, vasculitis, and a cellular variant of non-specific interstitial pneumonia. Another important condition associated with the RHS is pulmonary thromboembolism,^{46,49} where the diagnosis would be based on clinicoradiologic grounds. There are also case reports of association with other diseases such as lipid pneumonia, lymphomatoid granulomatosis, lymphocytic interstitial pneumonia, tuberous sclerosis complex, hypersensitivity pneumonia, and bronchoalveolar carcinoma.

Histopathologic Correlation of the RHS

The first pathologic description of the RHS was given by Voloudaki et al¹ in cryptogenic organizing pneumonia. The central ground-glass opacity was due to alveolar septal inflammation with intact alveolar air spaces. The denser rim was due to inflammation/granulation tissue within the air space itself. A similar pathologic description has also been provided in other reports.^{6,8,54} In all the aforementioned cases, the central ground-glass area had only alveolar septal inflammation, whereas the denser rim was composed of intra-alveolar infiltrate. However, in invasive fungal infections, the RHS is due to infarct, with greater hemorrhage at the periphery than in the center.²⁰ The central infarct may show multiple levels of pulmonary arteries and veins extensively occluded by thrombi containing numerous *Zygomycetes* hyphae. In the peripheral ring, inflammation, massive hemorrhage, and fibrinous exudates were noted.⁴⁵ In a patient with tumor emboli, the RHS lesion at autopsy was composed of central necrotic islands surrounded by a peripheral ring-like hemorrhagic band. The pulmonary vasculature and lymphatics near the hemorrhagic band were occluded by tumor emboli.⁵² In lymphomatoid granulomatosis, the central ground-glass opacity is due to central necrosis, whereas the peripheral rim is due to the angioinvasive nature of lymphomatoid granulomatosis.¹⁴ Thus, a similar computed tomography (CT) appearance of the RHS may occur due to differing pathological processes. In angioinvasive fungal infections (and possibly in pulmonary thromboembolism), the central ground-glass opacity is due to infarct with preserved air spaces, whereas in organizing pneumonia, the central ground-glass opacity is due to inflammation of the alveolar septa alone. The rim of consolidation is due to dense inflammation, granulation tissue, or hemorrhage within the alveolar air spaces.

Discussion

The CT appearance of central ground-glass opacity within a ring- or crescent-shaped rim of consolidation (Fig. 2) was first described by Voloudaki et al¹ in two patients with cryptogenic organizing pneumonia. Similar descrip-

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Table 1. Details of Publications Reporting the Reversed Halo Sign

| Reference | Country | Final Diagnosis | Patients (n) | Age (y) | Sex (Male/Female) | Basis for Diagnosis | Immune Status |
|--------------------------------|---------|---|--------------|---------|-------------------|--|-------------------|
| Voloudaki et al ¹ | Greece | COP | 2 | 50, 53 | 1/1 | Histopathology (SLB) | Immunocompetent |
| Marlow et al ³ | USA | Sarcoidosis | 1 | 43 | 0/1 | Histopathology (TBLB) | Immunocompetent |
| Zompatori et al ⁴ | Italy | COP | 1 | 58 | 0/1 | Histopathology (TBLB) | Immunocompetent |
| Kondo et al ⁵ | Japan | Secondary OP (minocycline-induced) | 1 | 39 | 0/1 | Histopathology (TBLB) | Immunocompetent |
| Fujii et al ⁶ | Japan | Sarcoidosis with secondary OP | 1 | 40 | 1/0 | Histopathology (SLB) | Immunocompetent |
| Choi et al ⁷ | Korea | Small vessel vasculitis | 1 | 41 | 0/1 | Histopathology (VATS lung biopsy) | Immunocompetent |
| Kim et al ² | Korea | COP | 6 | 20–79 | NA | Histopathology | Immunocompetent |
| Gasparetto et al ⁸ | Brazil | Pulmonary paracoccidiodomycosis | 15 | 20–58 | 13/2 | BAL, bronchial or transbronchial lung biopsy | NA |
| Hata and Uehara ⁹ | Japan | Secondary OP (drug-related: Sho-sheriyu-to, an herbal medicine) | 1 | 67 | 1/0 | Radiologic finding and response to steroids and withdrawal of drug | Immunocompetent |
| Soberón et al ¹⁰ | Spain | Organizing pneumonia: cryptogenic in 2 patients, secondary OP in 2 patients | 4 | 22–87 | NA | Histopathology | Immunocompetent |
| Agarwal et al ¹¹ | India | GPA | 1 | 28 | 1/0 | C-ANCA positivity and renal biopsy suggestive of crescentic glomerulonephritis | Immunocompetent |
| Ahuja et al ¹² | India | Tuberculosis | 1 | 15 | 1/0 | Acid fast bacilli positive on VATS lung biopsy | Immunocompetent |
| Arai et al ¹³ | Japan | COP | 1 | 56 | 0/1 | Histopathology (TBLB) | Immunocompetent |
| Benamore et al ¹⁴ | Canada | Lymphomatoid granulomatosis | 1 | 50 | 0/1 | Histopathology (SLB) | Immunocompetent |
| Hsu et al ¹⁵ | NA | Secondary OP (chronic lymphocytic leukemia) | 1 | 54 | 1/0 | Histopathology (needle aspiration) | Immunocompetent |
| Kanaji et al ¹⁶ | Japan | Exogenous lipid pneumonia | 1 | 50 | 1/0 | Histopathology (VATS lung biopsy) | Immunocompetent |
| Okubo et al ¹⁷ | Japan | Psittacosis | 1 | 52 | 0/1 | Elevated antibody against <i>Chlamydia psittaci</i> and response to minocycline | Immunocompetent |
| Sakai et al ¹⁸ | Japan | <i>Legionella pneumophila</i> pneumonia | 2 | 27–91 | NA | Microbiological evidence | NA |
| Ueda et al ¹⁹ | Japan | Idiopathic nonspecific interstitial pneumonia | 1 | 39 | 1/0 | Histopathology (VATS lung biopsy) | Immunocompetent |
| Wahba et al ²⁰ | Texas | Mucormycosis Invasive aspergillosis | 7 1 | 24–70 | 5/3 | BAL (4 patients) TTNA (2 patients) TBLB (1 patient) Lobectomy (1 patient) | Immunocompromised |
| Bräunlich et al ²¹ | Germany | Lymphomatoid granulomatosis | 1 | 79 | 1/0 | Histopathology (SLB) | Immunocompetent |
| Kumazoe et al ²² | Japan | Sarcoidosis | 1 | 27 | 1/0 | Histopathology (TBLB) | Immunocompetent |
| Tzouveleki et al ²³ | Greece | Acute fibrinous and OP | 1 | 65 | 0/1 | Histopathology (VATS lung biopsy) | Immunocompetent |
| Burke et al ²⁴ | UK | COP | 1 | 40 | 0/1 | Radiologic evidence and response to steroids | Immunocompetent |

(continued)

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Table 1. Details of Publications Reporting the Reversed Halo Sign (continued)

| Reference | Country | Final Diagnosis | Patients (n) | Age (y) | Sex (Male/Female) | Basis for Diagnosis | Immune Status |
|-----------------------------------|---------|---|--------------|---------|-------------------|--|---|
| Chung et al ²⁵ | USA | Pulmonary mucormycosis | 1 | 64 | 0/1 | Histopathology (CT-guided biopsy) | Immunocompromised (post-HSCT, diabetes) |
| Godbert et al ²⁶ | France | COP | 1 | 51 | 0/1 | Clinicoradiologic evidence and response to steroids | Immunocompetent |
| Ito et al ²⁷ | Japan | Secondary OP (bird fancier's lung) | 1 | 70 | 1/0 | Exposure to pigeons, antibody positivity, and histopathology (SLB) | Immunocompetent |
| Maimon ²⁸ | Canada | COP | 1 | 47 | 0/1 | Histopathology (SLB) | Immunocompetent |
| Marchiori et al ²⁹ | Brazil | Tuberculosis | 1 | 32 | 0/1 | Sputum smear positivity and histopathology (SLB) | Immunocompetent |
| Marchiori ³⁰ | Brazil | Sarcoidosis | 1 | 44 | 0/1 | Histopathology (SLB) | Immunocompetent |
| Otera et al ³¹ | Japan | <i>Pneumocystis</i> pneumonia | 1 | 32 | 1/0 | Microbiologic evidence on lung biopsy | Immunocompromised (AIDS) |
| Tokuyasu et al ³² | Japan | Secondary OP (dermatomyositis) | 1 | 55 | 0/1 | Histopathology (VATS lung biopsy) | Immunocompetent |
| Tzilas et al ³³ | Greece | Secondary OP (pneumococcal pneumonia) | 1 | 57 | 0/1 | Urinary antigen positivity and response to moxifloxacin | Immunocompetent |
| Walsh and Robertson ³⁴ | UK | COP | 1 | NA | 1/0 | Histopathology (SLB) | Immunocompetent |
| Anai et al ³⁵ | Japan | Secondary OP (Sjogren syndrome) | 1 | 41 | 0/1 | SS-A positivity and histopathology (TBLB and lip biopsy) | Immunocompetent |
| Godoy and Marom ³⁶ | USA | Pulmonary mucormycosis | 1 | 24 | 0/1 | Microbiologic evidence (TBLB) | Immunocompromised (acute myeloid leukemia chemotherapy) |
| Gudavalli et al ³⁷ | USA | Secondary OP (post-irradiation) | 2 | 65, 71 | 0/2 | Histopathology (TBLB) | Immunocompetent |
| Hong et al ³⁸ | Korea | Idiopathic nonspecific interstitial pneumonia | 1 | 52 | 0/1 | Histopathology (VATS lung biopsy) | Immunocompetent |
| Inoue et al ³⁹ | Japan | COP | 7 | 46-73 | 4/3 | Histopathology (TBLB) | Immunocompetent |
| Mango et al ⁴⁰ | USA | Post-RFA (likely secondary OP) | 1 | 80 | 1/0 | History of RFA and spontaneous resolution | Immunocompetent |
| Marchiori et al ⁴¹ | Brazil | Pulmonary histoplasmosis | 1 | 23 | 1/0 | Microbiologic evidence (BAL) | Immunocompetent |
| Mehta et al ⁴² | USA | COP | 1 | 54 | 0/1 | Histopathology (TBLB) | Immunocompetent |
| Mukai et al ⁴³ | Japan | Secondary OP (post-radiation) | 1 | 69 | 0/1 | Histopathology (TBLB) | Immunocompetent |
| Busca et al ⁴⁴ | Italy | Pulmonary mucormycosis | 1 | 55 | 0/1 | Microbiologic evidence (TBLB) | Immunocompromised (post-HSCT) |
| Kimura et al ⁴⁵ | Japan | <i>Cunninghamella</i> pneumonia | 1 | 53 | 0/1 | Histopathology (postmortem lung biopsy) | Immunocompromised (post-HSCT) |

(continued)

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Table 1. Details of Publications Reporting the Reversed Halo Sign (continued)

| Reference | Country | Final Diagnosis | Patients (n) | Age (y) | Sex (Male/Female) | Basis for Diagnosis | Immune Status |
|------------------------------------|---------|--|--------------|---------|-------------------|--|---|
| Marchiori et al ⁴⁶ | Brazil | Organizing pneumonia: 18; paracoccidioidomycosis: 12; aspergillosis: 6; mucormycosis: 6; tuberculosis: 12; cryptococcosis: 1; histoplasmosis: 1; pulmonary embolism: 7; sarcoidosis: 5; GPA: 2; pulmonary edema: 3; bronchoalveolar carcinoma: 3; <i>Pneumocystis jirovecii</i> pneumonia, 1 | 79 | 21–77 | 44/35 | Varied | NA |
| Markos et al ⁴⁷ | USA | Secondary OP (cutaneous T-cell lymphoma) | 1 | 53 | 0/1 | Histopathology (VATS lung biopsy) | Immunocompetent |
| Valente et al ⁴⁸ | Italy | H1N1 pneumonia and ARDS | 3 | 21–76 | NA | RT-PCR positivity | Immunocompetent |
| Casullo and Semionov ⁴⁹ | Canada | Pulmonary thromboembolism | 12 | 21–80 | 5/7 | Computed tomography pulmonary angiography (11 patients), multidetector-row computed tomography (1 patient) | Immunocompetent |
| Choo et al ⁵⁰ | Korea | Mucormycosis | 2 | 43, 47 | 2/0 | Histopathology | Immunosuppressed |
| Freeman et al ⁵¹ | USA | Lymphocytic interstitial pneumonia | 1 | 52 | 0/1 | Histopathology (CT-guided biopsy) | Immunocompetent |
| Goshima et al ⁵² | Japan | Pulmonary infarct (tumor emboli) | 1 | 79 | 0/1 | Postmortem lung biopsy | Immunocompetent |
| Madan and Guleria ⁵³ | India | COP | 1 | 14 | 0/1 | Histopathology (CT-guided biopsy) | Immunocompetent |
| Okubo et al ⁵⁴ | Japan | Mucormycosis | 1 | 64 | 1/0 | Histopathology (lobectomy) | Immunocompromised (acute lymphoblastic leukemia chemotherapy) |
| Stewart et al ⁵⁵ | USA | Mucormycosis | 1 | 63 | 0/1 | Histopathology (TBLB) | Immunocompromised (renal transplant) |
| Suzuki et al ⁵⁶ | Japan | Tuberous sclerosis complex | 1 | NA | 0/1 | CT-guided TTNA | Immunocompetent |
| Yüksekçaya et al ⁵⁷ | Turkey | Hypersensitivity pneumonitis | 1 | 54 | 1/0 | Histopathology (VATS lung biopsy) | Immunocompetent |
| Zhan et al ⁵⁸ | China | Tuberculosis | 17 | 36 ± 14 | 7/10 | Histopathology (CT-guided biopsy) | Immunocompetent |

COP = cryptogenic organizing pneumonia

SLB = surgical lung biopsy

TBLB = transbronchial lung biopsy

OP = organizing pneumonia

VATS = video-assisted thoracoscopic surgery

NA = not available

BAL = bronchoalveolar lavage

GPA = granulomatosis with polyangiitis

C-ANCA = anti-neutrophil cytoplasmic antibody

TTNA = transthoracic needle aspiration

CT = computed tomography

HSCT = hematopoietic stem cell transplant

AIDS = acquired immune deficiency syndrome

RFA = radiofrequency ablation

RT-PCR = reverse transcription polymerase chain reaction

Table 2. Disorders Manifesting With the Reversed Halo Sign on Computed Tomography of the Chest

| | No. |
|--|-----|
| Infectious diseases | |
| Pulmonary mucormycosis ^{20,25,36,44,45,50,54,55,65} | 23 |
| Invasive pulmonary aspergillosis ^{20,65} | 8 |
| Histoplasmosis ⁴¹ | 1 |
| Cryptococcosis ⁴⁶ | 1 |
| Paracoccidioidomycosis ^{8,46} | 27 |
| <i>Pneumocystis jirovecii</i> pneumonia ^{31,46} | 2 |
| H1N1 ARDS ⁴⁸ | 3 |
| Tuberculosis ^{12,46,58} | 30 |
| Psittacosis ¹⁷ | 1 |
| <i>Legionella pneumophila</i> pneumonia ¹⁸ | 2 |
| Noninfectious diseases | |
| Cryptogenic organizing pneumonia ^{1,2,4,10,13,23,24,26,28,34,39,53,65} | 36 |
| Secondary organizing pneumonia ^{5,6,9,10,15,27,32,33,35,37,40,43,47,65} | 30 |
| Vasculitis ^{7,11,46} | 4 |
| Sarcoidosis ^{3,22,46} | 7 |
| Lymphomatoid granulomatosis ^{14,21} | 2 |
| Idiopathic nonspecific interstitial pneumonia ^{19,38} | 2 |
| Lipoid pneumonia ¹⁶ | 1 |
| Pulmonary thromboembolism ^{46,49} | 19 |
| Pulmonary tumor embolus ⁵² | 1 |
| Pulmonary edema ⁴⁶ | 3 |
| Bronchoalveolar carcinoma ⁴⁶ | 3 |
| Hypersensitivity pneumonitis ⁵⁷ | 1 |
| Tuberous sclerosis complex ⁵⁶ | 1 |
| Lymphocytic interstitial pneumonia ⁵¹ | 1 |
| Total | 209 |

tions of such ring or annular opacities were also provided by Kondo et al⁵ and Fujii et al,⁶ who described this pattern in patients with secondary organizing pneumonia. The term “reversed halo sign” was first used by Moon et al⁵⁹ when they described four patients with cryptogenic organizing pneumonia with this radiologic sign. Subsequently, the same group compared the CT findings of 31 patients with biopsy-proven cryptogenic organizing pneumonia with 30 patients without cryptogenic organizing pneumonia. The RHS was noted in 6 patients with cryptogenic organizing pneumonia (19%) and in none with other diseases.² The authors concluded that the presence of the RHS is a specific radiologic finding of cryptogenic organizing pneumonia.

The RHS was initially defined by Kim et al² as a central ground-glass opacity surrounded by a denser consolidation with a crescentic (forming more than three fourths of a circle) or ring (forming a complete circle) shape of at least 2 mm in thickness. This description was slightly modified by the Fleischner Society in 2008,⁶⁰ which describes the RHS as a focal rounded area of ground-glass opacity surrounded by a more or less complete ring of consolidation.



Fig. 2. High-resolution computed tomography of the chest wall showing multiple areas with the reversed halo sign (arrows).

This CT appearance has been variously described by different authors as reversed halo sign, reverse halo sign, atoll sign, or fairy ring sign. Even before the term RHS was coined, Zompatori et al⁴ used the term “atoll sign” to describe the same CT finding in 1999. The word atoll is derived from atholhu, a Maldivian word meaning an island consisting of a lagoon surrounded by a circular coral reef.

In 1999, Marlow et al³ used the term “fairy ring sign” in a patient with sarcoidosis to describe a ring-shaped opacity with normal appearing lung parenchyma in the center. The term “fairy ring” is derived from Gaelic mythology. Fairies would dance in small circles at night, and when tired, they would rest on the toadstools. The next day, one would see only a circle in the grass with a ring of mushrooms, which is described as a fairy ring. In a strict sense, the fairy ring sign differs from the RHS in that the center of the ring is normal appearing lung parenchyma in the former and ground-glass opacity in the latter. However, some authors tend to use the terms interchangeably, as the difference between the two is only minor, and both signs are seen in the same diseases. On occasion, the same chest CT may show both the fairy ring sign and the RHS.³⁴ As the terms atoll sign and fairy ring sign are not mentioned in the Fleischner glossary of radiologic terms, we suggest using the term RHS to describe this CT finding.

It was suggested previously that in an immunocompetent host, the presence of an RHS would be sufficient evidence for a trial of steroids and obviate the need for a lung biopsy.²⁶ As shown in the systematic review, the RHS is associated with a number of diseases, both infectious and noninfectious. In fact, the presence of the RHS in an immunocompromised host should be attributed to invasive fungal infection unless proven otherwise. The RHS is also an early indicator of fungal infection, as it usually occurs within the first week of infection.⁶¹ In a study on

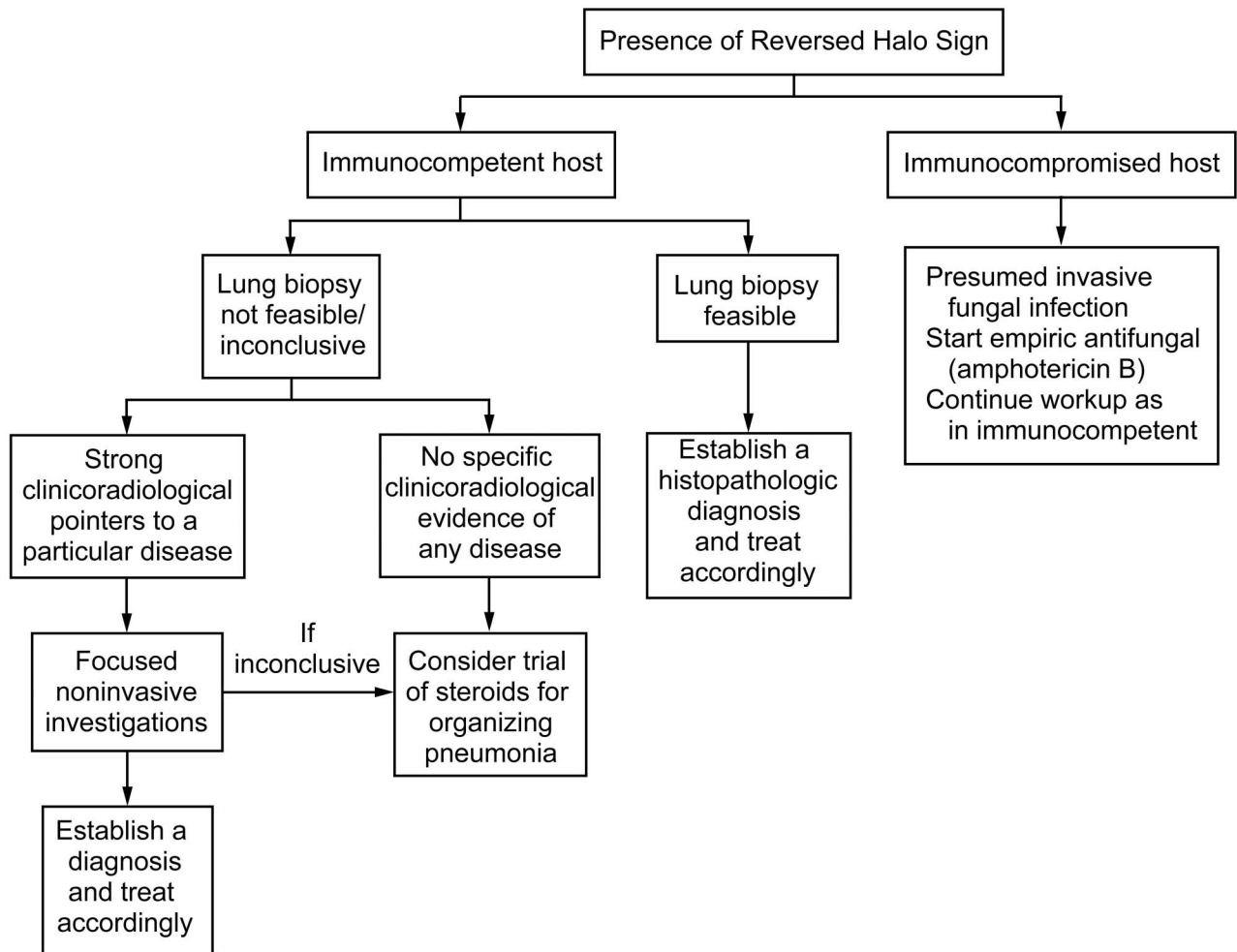


Fig. 3. Algorithmic approach for the reversed halo sign.

sequential morphological changes in CT scan in pulmonary mucormycosis, it was shown that the halo sign and the RHS are the initial radiologic features. With treatment, they develop areas of central cavitation.⁵⁰ As opposed to the halo sign, which is more common with invasive aspergillosis, the RHS is most commonly seen with mucormycosis, and thus, its presence would indicate the need for a *Mucor*-specific antifungal agent such as amphotericin B or posaconazole.⁶² Also, it would be appropriate to rule out endemic infections such as paracoccidioidomycosis and tuberculosis depending on the patient's geographic location by appropriate investigations in patients presenting with RHS. To date, the RHS has not been described in typical community-acquired bacterial pneumonia, and hence, the presence of this sign should rule out a diagnosis of community-acquired pneumonia. With the growing number of diseases with which it is being associated, especially infectious causes, we would not recommend an empiric steroid trial (as proposed earlier) without an attempt made to confirm the diagnosis.²⁶

Of the noninfectious causes of the RHS, the most commonly associated disease is organizing pneumonia. As the RHS can be encountered in both primary (cryptogenic organizing pneumonia) and secondary organizing pneumonia, it does not help in differentiating between cryptogenic and other causes. Thus, appropriate investigations should be carried out to rule out secondary causes (history of drugs, radiation exposure, connective tissue disease), which, if present, need to be treated simultaneously. Although many noninfectious diseases associated with the RHS (organizing pneumonia, hypersensitivity pneumonitis, sarcoidosis, vasculitis, cellular nonspecific interstitial pneumonia) might eventually need immunosuppression with steroids, it is important to confirm the diagnosis before starting therapy, as the degree and duration of immunosuppression needed in the individual disorders may vary. There are also reports of association with other rare diseases such as lipoid pneumonia, lymphomatoid granulomatosis, and bronchoalveolar carcinoma, further underscoring the need for a lung biopsy.

Marchiori et al⁶³⁻⁶⁵ have proposed several radiologic criteria to distinguish various diseases based upon the morphologic appearance of the RHS on the CT scan. They propose that the presence of nodular walls or nodules within the lesion indicates granulomatous diseases such as tuberculosis and sarcoidosis rather than organizing pneumonia.^{63,64} Similarly, a greater thickness of the rim (>1 cm), the presence of reticulation within the rim of consolidation, and the presence of an associated pleural effusion point toward a fungal pneumonia rather than organizing pneumonia.⁶⁵

Clinical Approach to a Case with RHS

As the RHS is not specific for any disorder, we recommend an algorithmic approach as outlined in Figure 3. The overall clinical presentation needs to be considered while approaching a patient with RHS. In an immunocompromised host, amphotericin B should be initiated empirically for presumed invasive fungal infection, and further investigations should proceed depending on the clinical circumstances. In an immunocompetent host, we recommend lung biopsy wherever feasible before initiating treatment. Flexible bronchoscopy with bronchoalveolar lavage and lung biopsy should be the initial investigation, with CT-guided or surgical lung biopsy as alternative options. However, if a lung biopsy is not feasible, is contraindicated, or is inconclusive, focused noninvasive investigations should be performed as guided by the clinicoradiologic presentation of the patient to achieve a diagnosis. In countries endemic for tuberculosis or paracoccidioidomycosis, appropriate investigations have to be carried out to rule out these infections. Other investigations such as anti-neutrophil cytoplasmic antibody may be useful in the diagnosis of vasculitides. It is important to make a conclusive diagnosis before initiating treatment, as this sign is associated with both infectious and noninfectious diseases, for which the treatment approaches are totally differ. If lung biopsy and noninvasive investigations do not help in reaching a diagnosis, a trial of empiric steroids may be started once infections have been reasonably excluded.

Finally, our review is not without limitations. Although a systematic review, the data presented are a collection of case reports and case series, and hence, we cannot exclude publication and reporting biases.

Conclusions

The RHS, previously considered to be a hallmark of cryptogenic organizing pneumonia, can be encountered in a variety of pulmonary disorders, both infectious and non-infectious. The two most commonly associated diseases are the organizing pneumonias and invasive fungal pneumonias (especially mucormycosis). As the treatment ap-

proach for these two diseases is diametrically opposite, we recommend obtaining a tissue diagnosis whenever feasible. The clinical presentation, immune status of the patient, and associated radiologic findings can help in narrowing down the differential diagnosis. In an immunocompromised host, antifungal therapy for mucormycosis should be initiated pending further investigations.

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