

Guidelines for Preventing Health-Care-Associated Pneumonia, 2003 Recommendations of the CDC and the Healthcare Infection Control Practices Advisory Committee

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Summary

This report updates, expands, and replaces the previously published Centers for Disease Control and Prevention (CDC) Guideline for Prevention of Nosocomial Pneumonia. The new guidelines are designed to reduce the incidence of pneumonia

The material in this report originated in the National Center for Infectious Diseases, Centers for Disease Control and Prevention, Atlanta, Georgia: James M Hughes MD, Division of Healthcare Quality Promotion, and Denise M Cardo MD, Director; and the Division of Bacterial and Mycotic Diseases, Mitchell L Cohen MD, Director.

This article is an abridgement of the CDC *Recommendations and Reports, Morbidity and Mortality Weekly Report*, March 26, 2004, *MMWR Recomm Rep* 2004;53(RR-3):1–36. This version of the report reflects corrections of errata in the original publication. Reprinted here are the Summary; Introduction; Key terms used in the guidelines; the section on Health-Care-Associated Bacterial Pneumonia from Part II—Recommendations and from Part III—Performance Indicators; and associated references. The complete CDC document with Parts I, II, and III is available online at <http://www.cdc.gov/ncidod/hip/pneumonia/default.htm>.

A full listing of the members of the Healthcare Infection Control Practices Committee is available online.

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and other severe, acute, lower-respiratory-tract infections in acute-care hospitals and in other health-care settings (eg, ambulatory and long-term care institutions) and other facilities where health care is provided.

Among the changes in the recommendations to prevent bacterial pneumonia, especially ventilator-associated pneumonia, are the preferential use of orotracheal rather than nasotracheal tubes in patients who receive mechanically assisted ventilation, the use of noninvasive ventilation to reduce the need for and duration of endotracheal intubation, changing the breathing circuits of ventilators when they malfunction or are visibly contaminated, and (when feasible) the use of an endotracheal tube with a dorsal lumen to allow drainage of respiratory secretions; no recommendations were made about the use of sucalfate, histamine-2 receptor antagonists, or antacids for stress-bleeding prophylaxis. [Editor's note: The sections described in the remainder of the Summary are not included in the following reprint of the guidelines.] For prevention of health-care-associated Legionnaires disease, the changes include maintaining potable hot water at temperatures not suitable for amplification of *Legionella* species, considering routine culturing of water samples from the potable

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water system of a facility's organ-transplant unit when it is done as part of the facility's comprehensive program to prevent and control health-care-associated Legionnaires disease, and initiating an investigation for the source of *Legionella* species when one definite or one possible case of laboratory-confirmed health-care-associated Legionnaires disease is identified in an in-patient hemopoietic stem-cell transplant (HSCT) recipient or in 2 or more HSCT recipients who had visited an out-patient HSCT unit during all or part of the 2–10-day period before illness onset. In the section on aspergillosis, the revised recommendations include the use of a room with high-efficiency particulate air filters rather than laminar airflow as the protective environment for allogeneic HSCT recipients, and the use of high-efficiency respiratory-protection devices (eg, N95 respirators) by severely immunocompromised patients when they leave their rooms when dust-generating activities are ongoing in the facility. In the respiratory syncytial virus (RSV) section, the new recommendation is to determine, on a case-by-case basis, whether to administer monoclonal antibody (palivizumab) to certain infants and children aged < 24 months who were born prematurely and are at high risk for RSV infection. In the section on influenza, the new recommendations include the addition of oseltamivir (to amantadine and rimantadine) for prophylaxis of all patients without influenza illness and oseltamivir and zanamivir (to amantadine and rimantadine) as treatment for patients who are acutely ill with influenza in a unit where an influenza outbreak is recognized.

In addition to the revised recommendations, the guidelines contain new sections on pertussis and lower respiratory tract infections caused by adenovirus and human parainfluenza viruses and refers readers to the source of updated information about prevention and control of severe acute respiratory syndrome.

Introduction

Because of the high morbidity and mortality associated with health-care-associated pneumonia, several guidelines for its prevention and control have been published. The first CDC Guideline for Prevention of Nosocomial Pneumonia was published in 1981 and addressed the main infection-control problems related to hospital-acquired pneumonia at the time: the use of large-volume nebulizers that were attached to mechanical ventilators and improper reprocessing (ie, cleaning and disinfection or sterilization) of respiratory-care equipment. The document also covered the prevention and control of hospital-acquired influenza and RSV infection.

In 1994 the Healthcare Infection Control Practices Advisory Committee (HICPAC) revised and expanded the CDC Guideline for Prevention of Nosocomial Pneumonia

to include Legionnaires disease and pulmonary aspergillosis.¹ HICPAC advises the Secretary of Health and Human Services, the directors of CDC, and the National Center for Infectious Diseases about the prevention and control of health-care-associated infections and related adverse events. The 1994 guideline addressed concerns related to preventing ventilator-associated pneumonia (VAP) (eg, the role of stress-ulcer prophylaxis in the causation of pneumonia and the contentious roles of selective gastrointestinal decontamination and periodic changes of ventilator tubing in the prevention of the infection). The report also presented major changes in the recommendations to prevent and control hospital-acquired pneumonia caused by *Legionella* species and aspergilli.

In recent years demand has increased for guidance on preventing and controlling pneumonia and other lower respiratory tract infections in health care settings other than the acute-care hospital, probably resulting in part from the progressive shift in the burden and focus of health care in the United States away from in-patient care in the acute-care hospital and toward out-patient and long-term care in other healthcare settings. In response to this, demand HICPAC revised the guideline to cover these other settings. However, infection-control data about the acute-care hospital setting are more abundant and well-analyzed; in comparison, data are limited from long-term care, ambulatory, and psychiatric facilities and other health-care settings.

This report consists of Parts II and III of a 3-part document² and contains the consensus HICPAC recommendations for the prevention of the following infections: bacterial pneumonia, Legionnaires disease, pertussis, invasive pulmonary aspergillosis (IPA), lower respiratory tract infections caused by RSV, parainfluenza and adenoviruses, and influenza. [Editor's note: This reprint consists of the sections on bacterial pneumonia *only*.) Part III provides suggested performance indicators to assist infection-control personnel in monitoring the implementation of the guideline recommendations in their facilities.

Part I of the guidelines [see <http://www.cdc.gov/ncidod/hip/pneumonia/default.htm>] provides the background for the recommendations and includes a discussion of the epidemiology, diagnosis, pathogenesis, modes of transmission, and prevention and control of the infections.² Part I can be an important resource for educating health care personnel. Because education of health care personnel is the cornerstone of an effective infection-control program, health care agencies should give high priority to continuing infection-control education programs for their staff members.

HICPAC recommendations address such issues as education of health care personnel about the prevention and control of health care-associated pneumonia and other lower respiratory tract infections, surveillance and reporting of

diagnosed cases of infections, prevention of person-to-person transmission of each disease, and reduction of host risk for infection.

Lower respiratory tract infection caused by *Mycobacterium tuberculosis* is not addressed in this document; however, it is covered in a separate publication.³

The document was prepared by CDC; reviewed by experts in infection control, intensive-care medicine, pulmonology, respiratory therapy, anesthesiology, internal medicine, and pediatrics; and approved by HICPAC. The recommendations are endorsed by the American College of Chest Physicians, American Healthcare Association, Association for Professionals of Infection Control and Epidemiology, Infectious Diseases Society of America, Society for Healthcare Epidemiology of America, and Society of Critical Care Medicine.

Key Terms Used in the Guidelines

Protective environment is a specialized patient-care area, usually in a hospital, with a positive air flow relative to the corridor (ie, air flows from the room to the outside adjacent space). The combination of high-efficiency particulate air (HEPA) filtration, high numbers (> 12) of air changes per hour (ACH), and minimal leakage of air into the room creates an environment that can safely accommodate patients who have received allogeneic HSCT.

Immunocompromised patients are those patients whose immune mechanisms are deficient because of immunologic disorders (eg, human immunodeficiency virus [HIV] infection, congenital immune deficiency syndrome, and chronic diseases [diabetes mellitus, cancer, emphysema, or cardiac failure), or immunosuppressive therapy (eg, radiation, cytotoxic chemotherapy, anti-rejection medication, and steroids). Immunocompromised patients who are identified as patients at high risk have the greatest risk for infection and include persons with severe neutropenia (ie, an absolute neutrophil count of < 500 cells/mL) for prolonged periods of time, recipients of allogeneic HSCT, and those who receive the most intensive chemotherapy (eg, patients with childhood acute myelogenous leukemia).

Part II—Recommendations of the Healthcare Infection Control Practices Advisory Committee [abridged]

Categorization of Recommendations

In this document each recommendation is categorized on the basis of existing scientific evidence, theoretical rationale, applicability, and potential economic impact. In addition, a new category accommodates recommendations that are made on the basis of existing national or state

health regulations. The following categorization scheme is applied in these guidelines:

Category IA. Strongly recommended for implementation and strongly supported by well-designed experimental, clinical, or epidemiologic studies.

Category IB. Strongly recommended for implementation and supported by certain clinical or epidemiologic studies and by strong theoretical rationale.

Category IC. Required for implementation, as mandated by federal or state regulation or standard.

Category II. Suggested for implementation and supported by suggestive clinical or epidemiologic studies or by strong theoretical rationale.

No recommendation; unresolved issue. Practices for which insufficient evidence or no consensus exists about efficacy.

Prevention of Health-Care-Associated Bacterial Pneumonia

I. Staff Education and Involvement in Infection Prevention

Educate health care workers about the epidemiology of, and infection-control procedures for, preventing health-care-associated bacterial pneumonia to ensure worker competency according to the worker's level of responsibility in the health care setting, and involve the workers in the implementation of interventions to prevent health-care-associated pneumonia by using performance-improvement tools and techniques (IA).⁴⁻¹¹

II. Infection and Microbiologic Surveillance

A. Conduct surveillance for bacterial pneumonia in intensive care unit (ICU) patients who are at high risk for health-care-related bacterial pneumonia (eg, patients with mechanically assisted ventilation or selected postoperative patients) to determine trends and help identify outbreaks and other potential infection-control problems.^{12,13} The use of the new National Nosocomial Infection Surveillance system's surveillance definition of pneumonia is recommended.¹⁴ Include data on the causative microorganisms and their antimicrobial susceptibility patterns.¹⁵ Express data as rates (eg, number of infected patients or infections per 100 ICU days or per 1,000 ventilator days) to facilitate intrahospital comparisons and trend determination.^{12,16,17} Link monitored rates and prevention efforts and return data to appropriate health care personnel (IB).¹⁸

B. In the absence of specific clinical, epidemiologic, or infection-control objectives, do not routinely perform surveillance cultures of patients or of

equipment or devices used for respiratory therapy, pulmonary-function testing, or delivery of inhalation anesthesia (II).¹⁹⁻²²

III. Prevention of Transmission of Microorganisms

A. Sterilization or Disinfection and Maintenance of Equipment and Devices

1. General measures
 - a. Thoroughly clean all equipment and devices to be sterilized or disinfected (IA).^{23,24}
 - b. Whenever possible, use steam sterilization (by autoclaving) or high-level disinfection by wet heat pasteurization at > 70°C for 30 min for reprocessing semicritical equipment or devices (ie, items that come into direct or indirect contact with mucous membranes of the lower respiratory tract) that are not sensitive to heat and moisture (Table 1). Use low-temperature sterilization methods (as approved by the Office of Device Evaluation, Center for Devices and Radiologic Health, Food and Drug Administration) for equipment or devices that are heat- or moisture-sensitive.²⁴⁻²⁸ After disinfection, proceed with appropriate rinsing, drying, and packaging, taking care not to contaminate the disinfected items in the process (IA).^{23,24}
 - c. Preferentially use sterile water for rinsing reusable semicritical respiratory equipment

and devices when rinsing is needed after they have been chemically disinfected. If this is not feasible, rinse the device with filtered water (ie, water that has been through a 0.2- μ filter) or tap water, and then rinse with isopropyl alcohol and dry with forced air or in a drying cabinet (IB).²⁴

- d. Adhere to provisions in Food and Drug Administration's enforcement document for single-use devices that are reprocessed by third parties (IC).^{24,29}
2. Mechanical ventilators

Do not routinely sterilize or disinfect the internal machinery of mechanical ventilators (II).
3. Breathing circuits, humidifiers, and heat-and-moisture exchangers (HMEs)
 - a. Breathing circuits with humidifiers
 - 1) Do not change routinely, on the basis of duration of use, the breathing circuit (ie, ventilator tubing and exhalation valve and the attached humidifier) that is in use on an individual patient. Change the circuit when it is visibly soiled or mechanically malfunctioning (IA).³⁰⁻³⁵
 - 2) Breathing-circuit-tubing condensate.
 - a) Periodically drain and discard any condensate that collects in the tubing of a mechanical ventilator, taking precautions not to allow condensate to drain toward the patient (IB).³⁶
 - b) Wear gloves to perform the previous procedure and/or when handling the fluid (IB).^{37,38}
 - c) Decontaminate hands with soap and water (if hands are visibly soiled) or with an alcohol-based hand rub after performing the procedure or handling the fluid (IA).^{38,39}
 - 3) No recommendation can be made for placing a filter or trap at the distal end of the expiratory-phase tubing of the breathing circuit to collect condensate (Unresolved issue).
 - 4) Humidifier fluids
 - a) Use sterile (not distilled, nonsterile) water to fill bubbling humidifiers (II).^{36,40-43}
 - b) No recommendation can be made for the preferential use of a closed, continuous-feed humidification system (Unresolved issue).
 - b. Ventilator breathing circuits with HMEs
 - 1) No recommendation can be made for the preferential use of either HMEs or heated

Table 1. Examples of Semicritical Items* Used on the Respiratory Tract

Anesthesia device or equipment including:
Face mask or tracheal tube
- Inspiratory and expiratory tubing
- Y-piece
- Reservoir bag
- Humidifier
Breathing circuits of mechanical ventilators
Bronchoscopes and their accessories, except for biopsy forceps and specimen brush†
Endotracheal and endobronchial tubes
Laryngoscope blades
Mouthpieces and tubing of pulmonary-function testing equipment
Nebulizers and their reservoirs
Oral and nasal airways
Probes of carbon dioxide analyzers, air-pressure monitors
Resuscitation bags
Stylets
Suction catheters
Temperature sensors

*Items that directly or indirectly contact mucous membranes of the respiratory tract should be sterilized or subjected to high-level disinfection before re-use.

†Considered critical items and should be sterilized before re-use.

- humidifiers to prevent pneumonia in patients receiving mechanically assisted ventilation (Unresolved issue) (IB).⁴⁴⁻⁴⁹
- 2) Changing HME
 - a) Change an HME that is in use on a patient when it malfunctions mechanically or becomes visibly soiled (II).
 - b) Do not routinely change more frequently than every 48 hours an HME that is in use on a patient (II).⁵⁰⁻⁵²
 - 3) Do not change routinely (in the absence of gross contamination or malfunction) the breathing circuit attached to an HME while it is in use on a patient (II).⁵³
4. Oxygen humidifiers
 - a. Follow manufacturers' instructions for use of oxygen humidifiers (II,IC).^{29,54-56}
 - b. Change the humidifier-tubing (including any nasal prongs or mask) that is in use on one patient when it malfunctions or becomes visibly contaminated (II).
 5. Small-volume medication nebulizers: in-line and hand-held nebulizers
 - a. Between treatments on the same patient, disinfect, rinse with sterile water, or air-dry small-volume in-line or hand-held medication nebulizers (IB).⁵⁷⁻⁵⁹
 - b. Use only sterile fluid for nebulization, and dispense the fluid into the nebulizer aseptically (IA).^{40-42,58,60-62}
 - c. Whenever possible, use aerosolized medications in single-dose vials. If multidose medication vials are used, follow manufacturers' instructions for handling, storing, and dispensing the medications (IB).^{60,62-67}
 6. Mist tents
 - a. Between uses on different patients, replace mist tents and their nebulizers, reservoirs, and tubing with those that have been subjected to sterilization or high-level disinfection (II).⁶⁸
 - b. No recommendation can be made about the frequency of routinely changing mist-tent nebulizers, reservoirs, and tubing while in use on one patient (Unresolved issue).
 - c. Subject mist-tent nebulizers, reservoirs, and tubing that are used on the same patient to daily low-level disinfection (eg, with 2% acetic acid) or pasteurization followed by air-drying (II).⁶⁹
 7. Other devices used in association with respiratory therapy
 - a. Respirometer and ventilator thermometer: between their uses on different patients, sterilize or subject to high-level disinfection portable respirometers and ventilator thermometers (IB).⁷⁰⁻⁷⁴
 - b. Resuscitation bags
 - 1) Between their uses on different patients, sterilize or subject to high-level disinfection reusable hand-powered resuscitation bags (IB).⁷⁵⁻⁷⁹
 - 2) No recommendation can be made about the frequency of changing hydrophobic filters placed on the connection port of resuscitation bags (Unresolved issue).
8. Anesthesia machines and breathing systems or patient circuits
 - a. Do not routinely sterilize or disinfect the internal machinery of anesthesia equipment (IB).⁸⁰
 - b. Between uses on different patients, clean reusable components of the breathing system or patient circuit (eg, tracheal tube or face mask, inspiratory and expiratory breathing tubing, Y-piece, reservoir bag, humidifier, and tubing), and then sterilize or subject them to high-level liquid chemical disinfection or pasteurization in accordance with the device manufacturers' instructions for their reprocessing (IB).^{24,26}
 - c. No recommendation can be made about the frequency of routinely cleaning and disinfecting unidirectional valves and carbon dioxide absorber chambers (Unresolved issue).⁸¹
 - d. Follow published guidelines or manufacturers' instructions about in-use maintenance, cleaning, and disinfection or sterilization of other components or attachments of the breathing system or patient circuit of anesthesia equipment (IB).^{82,83}
 - e. No recommendation can be made for placing a bacterial filter in the breathing system or patient circuit of anesthesia equipment (Unresolved issue).^{3,84-89}
 9. Pulmonary function testing equipment
 - a. Do not routinely sterilize or disinfect the internal machinery of pulmonary-function testing machines between uses on different patients (II).^{90,91}
 - b. Change the mouthpiece of a peak flow meter or the mouthpiece and filter of a spirometer between uses on different patients (II).^{91,92}
 10. Room-air "humidifiers" and faucet aerators
 - a. Do not use large-volume room-air humidifiers that create aerosols (eg, by venturi principle, ultrasound, or spinning disk, and thus

actually are nebulizers) unless they can be sterilized or subjected to high-level disinfection at least daily and filled only with sterile water (II).^{40,93,94}

b. Faucet aerators

- 1) No recommendation can be made about the removal of faucet aerators from areas for immunocompetent patients (see also section on Legionnaires Disease [not reprinted here], Part II, Section I-C-1-d) (Unresolved issue).
- 2) If *Legionella* species are detected in the water of a transplant unit and until *Legionella* species are no longer detected by culture, remove faucet aerators in the unit (see also section on Legionnaires Disease [not reprinted here], Part II, Section I-C-1-d) (II).⁹⁵

B. Prevention of Person-to-Person Transmission of Bacteria

1. Standard Precautions

- a. Hand hygiene: Decontaminate hands by washing them with either antimicrobial soap and water, or with nonantimicrobial soap and water (if hands are visibly dirty or contaminated with proteinaceous material or are soiled with blood or body fluids), or by using an alcohol-based antiseptic agent (eg, hand rub) if hands are not visibly soiled after contact with mucous membranes, respiratory secretions, or objects contaminated with respiratory secretions, whether or not gloves are worn. Decontaminate hands as described previously, before and after contact with a patient who has an endotracheal or tracheostomy tube in place, and before and after contact with any respiratory device that is used on the patient, whether or not gloves are worn (IA).^{37,39}

b. Gloving

- 1) Wear gloves for handling respiratory secretions or objects contaminated with respiratory secretions of any patient (IB).³⁷
- 2) Change gloves and decontaminate hands as described previously, between contacts with different patients; after handling respiratory secretions or objects contaminated with secretions from one patient and before contact with another patient, object, or environmental surface; and between contacts with a contaminated body site and the respiratory tract of, or respiratory device on, the same patient (IA).^{37,39,96-98}

c. Gowning

When soiling with respiratory secretions from a patient is anticipated, wear a gown and change it after soiling occurs and before providing care to another patient (IB).^{37,97}

2. Care of patients with tracheostomy

- a. Perform tracheostomy under aseptic conditions (II).
- b. When changing a tracheostomy tube, wear a gown, use aseptic technique, and replace the tube with one that has undergone sterilization or high-level disinfection (IB).^{23,24,37}
- c. No recommendation can be made for the daily application of topical antimicrobial agent(s) at the tracheostoma (Unresolved issue).⁹⁹

3. Suctioning of respiratory tract secretions

(See also Section IV-B-1-d).

- a. No recommendation can be made for the preferential use of either the multiuse closed-system suction catheter or the single-use open-system suction catheter for prevention of pneumonia (Unresolved issue).^{44,100-102}
- b. No recommendation can be made about wearing sterile rather than clean gloves when performing endotracheal suctioning (Unresolved issue).
- c. No recommendation can be made about the frequency of routinely changing the in-line suction catheter of a closed-suction system in use on one patient (Unresolved issue).¹⁰³
- d. If the open-system suction is employed, use a sterile, single-use catheter (II).
- e. Use only sterile fluid to remove secretions from the suction catheter if the catheter is to be used for re-entry into the patient's lower respiratory tract (II).

IV. Modifying Host Risk for Infection

A. Increasing Host Defense Against Infection: Administration of Immune Modulators

1. Pneumococcal vaccination. Vaccinate patients at high risk for severe pneumococcal infections.
 - a. Administer the 23-valent pneumococcal polysaccharide vaccine to persons aged ≥ 65 years; persons aged 5-64 years who have chronic cardiovascular disease (eg, congestive heart failure or cardiomyopathy), chronic pulmonary disease (eg, chronic obstructive pulmonary disease [COPD] or emphysema, but not asthma), diabetes mellitus, alcoholism, chronic liver disease (eg, cirrhosis), or

cerebrospinal fluid leaks; persons aged 5–64 years who have functional or anatomic asplenia; persons aged 5–64 years who are living in special environments or social settings; immunocompromised persons aged \geq 5 years with human immunodeficiency virus (HIV) infection, leukemia, lymphoma, Hodgkin disease, multiple myeloma, generalized malignancy, chronic renal failure, nephrotic syndrome, or other conditions associated with immunosuppression (eg, receipt of HSCT, solid-organ transplant, or immunosuppressive chemotherapy, including long-term systemic corticosteroids); and persons in long-term-care facilities (IA).^{104–109}

- b. Administer the 7-valent pneumococcal polysaccharide protein-conjugate vaccine to all children aged $<$ 2 years and to children aged 24–59 months who are at increased risk for pneumococcal disease (eg, children with sickle-cell disease or other hemoglobinopathies, or children who are functionally or anatomically asplenic; children with HIV infection; children who have chronic disease, including chronic cardiac or pulmonary disease [except asthma], diabetes mellitus, or cerebrospinal fluid leak; and children with immunocompromising conditions, including malignancies, chronic renal failure or nephrotic syndrome, or receipt of immunosuppressive chemotherapy, including long-term corticosteroids, and receipt of solid-organ transplant). Consider administering the vaccine to children aged 24–59 months, with priority to children aged 24–35 months, children who are American Indians/Alaska Natives or black, and children who attend group child-care centers (IB).¹⁰⁴
 - c. In nursing homes and other long-term-care facilities, establish a standing-order program for the administration of 23-valent vaccine to persons at high risk for acquiring severe pneumococcal infections, including pneumococcal pneumonia (IA).^{105,110,111}
2. No recommendation can be made for the routine administration of preparations of granulocyte-colony stimulating factor or intravenous gamma globulin for prophylaxis against health-

care-associated pneumonia (Unresolved issue).^{112–117}

3. No recommendation can be made for the routine enteral administration of glutamine for prevention of health-care-associated pneumonia (Unresolved issue).^{118,119}

B. Precautions for Prevention of Aspiration

As soon as the clinical indications for their use are resolved, remove devices such as endotracheal, tracheostomy, and/or enteral (ie, oro- or nasogastric or jejunal) tubes from patients (IB).^{120–125}

1. Prevention of aspiration associated with endotracheal intubation
 - a. Use of noninvasive ventilation to reduce the need for and duration of endotracheal intubation.
 - 1) When feasible and not medically contraindicated, use noninvasive positive-pressure ventilation delivered continuously by face or nose mask, instead of performing endotracheal intubation in patients who are in respiratory failure and are not needing immediate intubation (eg, those who are in hypercapnic respiratory failure secondary to exacerbation of COPD or cardiogenic pulmonary edema) (II).^{126–129}
 - 2) When feasible and not medically contraindicated, use noninvasive ventilation as part of the weaning process (from mechanically assisted ventilation) to shorten the period of endotracheal intubation (II).¹³⁰
 - b. As much as possible, avoid repeat endotracheal intubation in patients who have received mechanically assisted ventilation (II).¹³¹
 - c. Unless contraindicated by the patient's condition, perform orotracheal rather than nasotracheal intubation on patients (IB).^{44,132,133}
 - d. If feasible, use an endotracheal tube with a dorsal lumen above the endotracheal cuff to allow drainage (by continuous or frequent intermittent suctioning) of tracheal secretions that accumulate in the patient's subglottic area (II).^{44,134–137}
 - e. Before deflating the cuff of an endotracheal tube in preparation for tube removal, or before moving the tube, ensure that secretions are cleared from above the tube cuff (II).
2. Prevention of aspiration associated with enteral feeding
 - a. In the absence of medical contraindication(s), elevate at an angle of 30–45 degrees

- the head of the bed of a patient at high risk for aspiration (eg, a person receiving mechanically assisted ventilation and/or who has an enteral tube in place) (II).¹³⁸⁻¹⁴⁰
- b. Routinely verify appropriate placement of the feeding tube (IB).¹⁴¹⁻¹⁴³
 - c. No recommendation can be made for the preferential use of small-bore tubes for enteral feeding (Unresolved issue).¹⁴⁴
 - d. No recommendation can be made for preferentially administering enteral feedings continuously or intermittently (Unresolved issue).¹⁴⁵⁻¹⁴⁸
 - e. No recommendation can be made for preferentially placing the feeding tubes, (eg, jejunal tubes) distal to the pylorus (Unresolved issue).¹⁴⁹⁻¹⁵⁵
3. Prevention or modulation of oropharyngeal colonization
 - a. Oropharyngeal cleaning and decontamination with an antiseptic agent: develop and implement a comprehensive oral hygiene program (that might include the use of an antiseptic agent) for patients in acute-care settings or residents in long-term-care facilities who are at high risk for health-care-associated pneumonia (II).^{156,157}
 - b. Chlorhexidine oral rinse
 - 1) No recommendation can be made for the routine use of an oral chlorhexidine rinse for the prevention of health-care-associated pneumonia in all postoperative or critically ill patients and/or other patients at high risk for pneumonia (Unresolved issue) (II).¹⁵⁸
 - 2) Use an oral chlorhexidine gluconate (0.12%) rinse during the perioperative period on adult patients who undergo cardiac surgery (II).¹⁵⁸
 - c. Oral decontamination with topical antimicrobial agents
 - 1) No recommendation can be made for the routine use of topical antimicrobial agents for oral decontamination to prevent VAP (Unresolved issue).¹⁵⁹
 4. Prevention of gastric colonization
 - a. No recommendation can be made for the preferential use of sucralfate, H₂-antagonists, and/or antacids for stress-bleeding prophylaxis in patients receiving mechanically assisted ventilation (Unresolved issue).¹⁶⁰⁻¹⁶⁷
 - b. No recommendation can be made for the routine selective decontamination of the digestive tract of all critically ill, mechani-

cally ventilated, or ICU patients (Unresolved issue).¹⁶⁸⁻²⁰⁰

- c. No recommendation can be made for routinely acidifying gastric feeding (Unresolved issue).^{201,202}

C. Prevention of Postoperative Pneumonia

1. Instruct preoperative patients, especially those at high risk for contracting pneumonia, about taking deep breaths and ambulating as soon as medically indicated in the postoperative period. Patients at high risk include those who will have abdominal aortic aneurysm repair, thoracic surgery, or emergency surgery; those who will receive general anesthesia; those who are aged ≥ 60 years; those with totally dependent functional status; those who have had a weight loss $> 10\%$; those using steroids for chronic conditions; those with recent history of alcohol use, history of COPD, or smoking during the preceding year; those with impaired sensorium, a history of cerebrovascular accident with residual neurologic deficit, or low (< 8 mg/dL) or high (> 22 mg/dL) blood urea nitrogen level; and those who will have received > 4 units of blood before surgery (IB).²⁰³⁻²⁰⁶
2. Encourage all postoperative patients to take deep breaths, move about the bed, and ambulate unless medically contraindicated (IB).²⁰⁵⁻²⁰⁷
3. Use incentive spirometry on postoperative patients at high risk for pneumonia (IB).²⁰⁵⁻²⁰⁷
4. No recommendation can be made about the routine use of chest physiotherapy on all postoperative patients at high risk for pneumonia (Unresolved issue).²⁰⁵⁻²⁰⁷

D. Other Prophylactic Procedures for Pneumonia

1. Administration of antimicrobial agents other than in selective decontamination of the digestive tract
 - a. Systemic antimicrobial prophylaxis.

No recommendation can be made about the routine administration of systemic antimicrobial agent(s) to prevent pneumonia in critically ill patients or in those receiving mechanically-assisted ventilation (Unresolved issue).^{200,208}
 - b. Scheduled changes in the class of antimicrobial agents used for empiric therapy

No recommendation can be made for scheduled changes in the class of antimicrobial agents used routinely for empiric treatment of suspected infections in a particular group of patients (Unresolved issue).^{209,210}

- Turning or rotational therapy
No recommendation can be made for the routine use of turning or rotational therapy, either by “kinetic” therapy or by continuous lateral rotational therapy (ie, placing patients on beds that turn on their longitudinal axes intermittently or continuously) for prevention of health-care-associated pneumonia in critically ill and immobilized patients (Unresolved issue).^{44,211–216}

[The remainder of Part II is not reprinted here. See <http://www.cdc.gov/ncidod/hip/pneumonia/default.htm>]

Part III—Performance Indicator [abridged]

To assist infection-control personnel in assessing personnel adherence to the recommendations, the following performance measure is suggested:

- Monitor rates of VAP; can use established benchmarks and definitions of pneumonia (eg, National Nosocomial Infection Surveillance definitions and rates).¹⁴ Provide feedback to the staff about the facility’s VAP rates and reminders about the need for personnel to adhere to infection-control practices that reduce the incidence of VAP.

REFERENCES

- CDC. Guideline for prevention of nosocomial pneumonia. *MMWR* 1997;46(No. RR-1).
- CDC. Guidelines for preventing health-care-associated pneumonia, 2003. Atlanta, GA: US Department of Health and Human Services, CDC, 2004. Available at <http://www.cdc.gov/ncidod/hip/pneumonia/default.htm>.
- CDC. Guidelines for preventing the transmission of tuberculosis in health-care facilities, 1994. *MMWR* 1994;43(No. RR-13).
- Brooks K, Whitten S, Quigley D. Reducing the incidence of ventilator-related pneumonia. *J Health Qual* 1998;20:14–19.
- Halm EA, Atlas SJ, Borowsky LH, et al. Understanding physician adherence with a pneumonia practice guideline: effects of patient, system, and physician factors. *Arch Intern Med* 2000;160:98–104.
- Katz DA. Barriers between guidelines and improved patient care: an analysis of AHCPR’s Unstable Angina Clinical Practice Guideline. *Health Serv Res* 1999;34:377–89.
- Kaye J, Ashline V, Erickson D, et al. Critical care bug team: a multidisciplinary team approach to reducing ventilator-associated pneumonia. *Am J Infect Control* 2000;28:197–201.
- Kelleghan SI, Salemi C, Padilla S, et al. An effective continuous quality improvement approach to the prevention of ventilator-associated pneumonia. *Am J Infect Control* 1993;21:322–30.
- Joiner GA, Salisbury D, Bollin GE. Utilizing quality assurance as a tool for reducing the risk of nosocomial ventilator-associated pneumonia. *Am J Med Qual* 1996;11:100–3.
- Nicotra D, Ulrich C. Process improvement plan for the reduction of nosocomial pneumonia in patients on ventilators. *J Nurs Care Qual* 1996;10:18–23.
- Zack JE, Garrison T, Trovillion E, et al. Effect of an education program aimed at reducing the occurrence of ventilator-associated pneumonia. *Crit Care Med* 2002;30:2407–12.
- Haley RW, Culver DH, White JW, et al. The efficacy of infection surveillance and control programs in preventing nosocomial infections in US hospitals. *Am J Epidemiol* 1985;121:182–205.
- Haley RW, Morgan WM, Culver DH, et al. Hospital infection control: recent progress and opportunities under prospective payment. *Am J Infect Control* 1985;13:97–108.
- CDC. NNIS criteria for determining nosocomial pneumonia. Atlanta, GA: US Department of Health and Human Services, CDC, 2003.
- Horan TC, White JW, Jarvis WR, et al. Nosocomial infection surveillance, 1984. *MMWR* 1986;35(SS-1):17–29.
- Gaynes RP, Solomon S. Improving hospital-acquired infection rates: the CDC experience. *Jt Comm J Qual Improv* 1996;22:457–67.
- Josephson A, Karanfil L, Alonso H, Watson A, Blight J. Risk-specific nosocomial infection rates. *Am J Med* 1991;91:131–7.
- Gaynes R, Richards C, Edwards J, et al. Feeding back surveillance data to prevent hospital-acquired infections. *Emerging Infect Dis* 2001;7:295–8.
- American Hospital Association Committee on Infection within Hospitals. Statement on microbiologic sampling in the hospital. *Hospitals* 1974;48:125–6.
- Eickhoff TC. Microbiologic sampling. *Hospitals* 1970;44:86–7.
- Finelli L, Livengood JR, Saiman L. Surveillance of pharyngeal colonization: detection and control of serious bacterial illness in low birth weight infants. *Pediatr Infect Dis J* 1994;13:854–9.
- Glupeczynski Y. Usefulness of bacteriologic surveillance cultures for monitoring infection in hospitalized patients. *Acta Clin Belg* 2001;56:38–45.
- Favero MS, Bond WW. Clinical disinfection of medical and surgical materials. In: Block S, ed. *Disinfection, sterilization, and preservation*. Philadelphia, PA: Lea and Febiger, 1991.
- Rutala WA, Weber DJ, Healthcare Infection Control Practices Advisory Committee: guideline for disinfection and sterilization in healthcare facilities. *MMWR* (in press).
- Cefai C, Richards J, Gould FK, McPeake P. An outbreak of *Acinetobacter* respiratory tract infection resulting from incomplete disinfection of ventilatory equipment. *J Hosp Infect* 1990;15:177–82.
- Craig DB, Cowan SA, Forsyth W, Parker SE. Disinfection of anesthesia equipment by a mechanized pasteurization method. *Can Anaesth Soc J* 1975;22:219–23.
- McDonald WL, Welch HJ, Keet JE. Antisepsis of endotracheal tubes and face masks. *Anesthesiology* 1955;16:206–13.
- Spaulding EH. Studies on the chemical sterilization of surgical instruments. *Surg Gynecol Obstet* 1939;69:738–44.
- Food and Drug Administration. Enforcement priorities for single-use devices reprocessed by third parties and hospitals. Rockville, MD: US DHHS, FDA, 2000.
- Dreyfuss D, Djedaini K, Weber P, et al. Prospective study of nosocomial pneumonia and of patient and circuit colonization during mechanical ventilation with circuit changes every 48 hours versus no change. *Am Rev Respir Dis* 1991;143:738–43.
- Fink JB, Krause SA, Barrett L, Schaaff D, Alex CG. Extending ventilator circuit change interval beyond 2 days reduces the likelihood of ventilator-associated pneumonia. *Chest* 1998;113:405–11.
- Hess D, Burns E, Romagnoli D, Kacmarek RM. Weekly ventilator circuit changes: a strategy to reduce costs without affecting pneumonia rates. *Anesthesiology* 1995;82:903–11.
- Kollef MH, Shapiro D, Fraser VJ, et al. Mechanical ventilation with or without 7-day circuit changes: a randomized controlled trial. *Ann Intern Med* 1995;123:168–74.

34. Kotilainen HR, Keroack MA. Cost analysis and clinical impact of weekly ventilator circuit changes in patients in intensive care unit. *Am J Infect Control* 1997;25:117-20.
35. Long MN, Wickstrom G, Grimes A, Benton CF, Belcher B, Stamm AM. Prospective, randomised study of ventilator-associated pneumonia in patients with one versus three ventilator circuit changes per week. *Infect Control Hosp Epidemiol* 1996;17:14-19.
36. Craven DE, Goularte TA, Make BJ. Contaminated condensate in mechanical ventilator circuits: a risk factor for nosocomial pneumonia? *Am Rev Respir Dis* 1984;129:625-8.
37. Garner JS. Guideline for isolation precautions in hospitals: the Hospital Infection Control Practices Advisory Committee. *Infect Control Hosp Epidemiol* 1996;17:53-80.
38. Gorman LJ, Sanai L, Notman AW, Grant IS, Masterton RG. Cross infection in an intensive care unit by *Klebsiella pneumoniae* from ventilator condensate. *J Hosp Infect* 1993;23:27-34.
39. CDC. Guideline for hand hygiene in health-care settings. *MMWR* 2002;51(No. RR-16).
40. Arnow PM, Chou T, Weil D, Shapiro EN, Kretzschmar C. Nosocomial Legionnaires' disease caused by aerosolized tap water from respiratory devices. *J Infect Dis* 1982;146:460-7.
41. Carson LA, Favero MS, Bond WW, Petersen NJ. Morphological, biochemical and growth characteristics of *Pseudomonas cepacia* from distilled water. *Appl Microbiol* 1973;25:476-83.
42. Favero MS, Carson LA, Bond WW. *Pseudomonas aeruginosa*: growth in distilled water from hospitals. *Science* 1971;173:836-8.
43. Rhame FS, Streifel A, McComb C, Boyle M. Bubbling humidifiers produce microaerosols which can carry bacteria. *Infect Control* 1986;7:403-7.
44. Cook D, De Jonghe B, Brochard L, Brun-Buisson C. Influence of airway management on ventilator-associated pneumonia: evidence from randomized trials. *JAMA* 1998;279:781-7.
45. Dreyfuss D, Djedaini K, Gros I, et al. Mechanical ventilation with heated humidifiers or heat and moisture exchangers: effects on patient colonization and incidence of nosocomial pneumonia. *Am J Respir Crit Care Med* 1995;151:986-92.
46. Hurni JM, Feihl F, Lazor R, Leuenberger P, Perret C. Safety of combined heat and moisture exchanger filters in long-term mechanical ventilation. *Chest* 1997;111:686-91.
47. Kirton OC, DeHaven B, Morgan J, Morejon O, Civetta J. A prospective, randomised comparison of an in-line heat moisture exchange filter and heated wire humidifiers: rates of ventilator-associated early-onset (community-acquired) or late-onset (hospital-acquired) pneumonia and incidence of endotracheal tube occlusion. *Chest* 1997;112:1055-9.
48. Roustan JP, Kienlen J, Aubas P, Aubas S, du Cailar J. Comparison of hydrophobic heat and moisture exchangers with heated humidifier during prolonged mechanical ventilation. *Intensive Care Med* 1992;18:97-100.
49. Thomachot L, Viviani X, Arnaud S, Boisson C, Martin CD. Comparing two heat and moisture exchangers, one hydrophobic and one hygroscopic, on humidifying efficacy and the rate of nosocomial pneumonia. *Chest* 1998;114:1383-9.
50. Boisson C, Viviani X, Arnaud S, Thomachot L, Miliani Y, Martin C. Changing a hydrophobic heat and moisture exchanger after 48 hours rather than 24 hours: a clinical and microbiologic evaluation. *Intensive Care Med* 1999;25:1237-43.
51. Daumal F, Colpart E, Manoury B, Mariani M, Daumal M. Changing heat and moisture exchangers every 48 hours does not increase the incidence of nosocomial pneumonia. *Infect Control Hosp Epidemiol* 1999;20:347-9.
52. Thomachot L, Viallet R, Viguier JM, Sidier B, Roulier P, Martin C. Efficacy of heat and moisture exchangers after changing every 48 hours rather than 24 hours. *Crit Care Med* 1998;26:477-81.
53. Salemi C, Padilla S, Canola T, Reynolds D. Heat-and-moisture exchangers used with biweekly circuit tubing changes: effect on costs and pneumonia rates. *Infect Control Hosp Epidemiol* 2000;21:737-9.
54. Golar SD, Sutherland LLA, Ford GT. Multipatient use of prefilled disposable oxygen humidifiers for up to 30 days: patient safety and cost analysis. *Respir Care* 1993;38:343-7.
55. Henderson E, Ledgerwood D, Hope KM, et al. Prolonged and multipatient use of prefilled disposable oxygen humidifier bottles: safety and cost. *Infect Control Hosp Epidemiol* 1993;14:463-8.
56. Seto WH, Ching TY, Yuen KY, Lam WK. Evaluating the sterility of disposable wall oxygen humidifiers, during and between use on patients. *Infect Control* 1990;11:604-5.
57. Craven DE, Lichtenberg DA, Goularte TA, Make BJ, McCabe WR. Contaminated medication nebulizers in mechanical ventilation circuits. Source of bacterial aerosols. *Am J Med* 1984;77:834-8.
58. Mastro TD, Fields BS, Breiman RF, Campbell J, Plikaytis BD, Spika JS. Nosocomial Legionnaires' disease and use of medication nebulizers. *J Infect Dis* 1991;163:667-71.
59. Reboli AC, Koshinski R, Arias K, Marks-Austin K, Stieritz D, Stull TL. An outbreak of *Burkholderia cepacia* lower respiratory tract infection associated with contaminated albuterol nebulization solution. *Infect Control Hosp Epidemiol* 1996;17:741-3.
60. Mertz JJ, Scharer L, McClement JH. A hospital outbreak of *Klebsiella pneumoniae* from inhalation therapy with contaminated aerosol solutions. *Am Rev Respir Dis* 1967;95:454-60.
61. Moffet HL, Williams T. Bacteria recovered from distilled water and inhalation therapy equipment. *Am J Dis Child* 1967;114:7-12.
62. Sanders CV Jr., Luby JP, Johanson WG Jr., Barnett JA, Sanford JP. *Serratia marcescens* infections from inhalation therapy medications: nosocomial outbreak. *Ann Intern Med* 1970;73:15-21.
63. Hamill RJ, Houston ED, Georgiou PR, et al. An outbreak of *Burkholderia* (formerly *Pseudomonas*) *cepacia* respiratory tract colonization and infection associated with nebulized albuterol therapy. *Ann Intern Med* 1995;122:762-6.
64. Harbarth S, Sudre P, Dharan S, Cadenas M, Pittet D. Outbreak of *Enterobacter cloacae* related to understaffing, overcrowding, and poor hygiene practices. *Infect Control Hosp Epidemiol* 1999;20:598-603.
65. Longfield R, Longfield J, Smith LP, Hyams KC, Strohmmer ME. Multidose medication vial sterility: an in-use study and a review of literature. *Infect Control* 1984;5:165-9.
66. Ramsey AH, Skonieczny P, Coolidge DT, Kurzynski TA, Proctor ME, Davis JP. *Burkholderia cepacia* lower respiratory tract infection associated with exposure to a respiratory therapist. *Infect Control Hosp Epidemiol* 2001;22:423-6.
67. Sheth NK, Post GT, Wisniewski TR, Uttech BV. Multi-dose vials versus single-dose vials: a study in sterility and cost-effectiveness. *J Clin Microbiol* 1983;17:377-9.
68. Moffet HL, Allan D. Survival and dissemination of bacteria in nebulizers and incubators. *Am J Dis Child* 1967;114:13-20.
69. Jakobsson B, Hjelte L, Nystrom B. Low level of bacterial contamination of mist tents used in home treatment of cystic fibrosis patients. *J Hosp Infect* 2000;44:37-41.
70. Cunha BA, Klimek JJ, Gracewski J, McLaughlin JC, Quintiliani R. A common source outbreak of *Acinetobacter* pulmonary infection traced to Wright respirometers. *Postgrad Med J* 1980;56:169-72.
71. Irwin RS, Demers RR, Pratter MR, et al. An outbreak of *Acinetobacter* infection associated with the use of a ventilator spirometer. *Respir Care* 1980;25:232-7.
72. Kaul R, Burt JA, Cork L, et al. Investigation of a multiyear multiple critical care unit outbreak due to relatively drug-sensitive *Acinetobacter baumannii*: risk factors and attributable mortality. *J Infect Dis* 1996;174:1279-87.

73. Rogues AM, Maugein J, Allery A, et al. Electronic ventilator temperature sensors as a potential source of respiratory tract colonization with *Stenotrophomonas maltophilia*. *J Hosp Infect* 2001;49:289–92.
74. Weems JJ, Jr. Nosocomial outbreak of *Pseudomonas cepacia* associated with contamination of reusable electronic ventilator temperature probes. *Infect Control Hosp Epidemiol* 1993;14:583–6.
75. Fierer J, Taylor PM, Gezon HM. *Pseudomonas aeruginosa* epidemic traced to delivery-room resuscitators. *N Engl J Med* 1967;276:991–6.
76. Stone JW, Das BC. Investigation of an outbreak of infection with *Acinetobacter calcoaceticus* in a special care baby unit. *J Hosp Infect* 1986;7:42–8.
77. Thompson AC, Wilder BJ, Powner DJ. Bedside resuscitation bags: a source of bacterial contamination. *Infect Control* 1985;6:231–2.
78. Weber DJ, Wilson MB, Rutala WA, Thomann CA. Manual ventilation bags as a source for bacterial colonization of intubated patients. *Am Rev Respir Dis* 1990;142:892–4.
79. Van Der Zwet WC, Parlevliet GA, Savelkoul PH, et al. Outbreak of *Bacillus cereus* infections in a neonatal intensive care unit traced to balloons used in manual ventilation. *J Clin Microbiol* 2000;38:4131–6.
80. Du Moulin GC, Sauberman AJ. The anesthesia machine and circle system are not likely to be sources of bacterial contamination. *Anesthesiology* 1977;47:353–8.
81. Bengtson JP, Brandberg A, Brinkhoff B, Sonander H, Stenqvist O. Low-flow anesthesia does not increase the risk of microbial contamination through the circle absorber system. *Acta Anaesth Scand* 1989;33:89–92.
82. American Association of Nurse Anesthetists. Infection control guide. 2nd ed, 1993. Chicago, Illinois, 1993.
83. American Society for Anesthesiologists. Prevention of nosocomial infections in patients: recommendations for Infection Control for the Practice of Anesthesiology. Park Ridge, Illinois: American Society of Anesthesiologists, 1991.
84. Berry AJ, Nolte FS. An alternative strategy for infection control of anesthesia breathing circuits: a laboratory assessment of the Pall HME Filter. *Anesth Analg* 1991;72:651–5.
85. Feeley TW, Hamilton WK, Xavier B, Moyers J, Eger EI. Sterile anesthesia breathing circuits do not prevent postoperative pulmonary infection. *Anesthesiology* 1981;54:369–72.
86. Garibaldi RA, Britt MR, Webster C, Pace NL. Failure of bacterial filters to reduce the incidence of pneumonia after inhalation anesthesia. *Anesthesiology* 1981;54:364–8.
87. Luttrupp HH, Berntman L. Bacterial filters protect anaesthetic equipment in a low-flow system. *Anaesthesia* 1993;48:520–3.
88. Ping FC, Oulton JL, Smith JA, Skidmore AG, Jenkins LC. Bacterial filters—are they necessary on anesthetic machines? *Anaesth Soc J* 1979;26:415–9.
89. Vezina DP, Trepanier CA, Lessard MR, Gourdeau M, Tremblay C. Anesthesia breathing circuits protected by the DAR Barrierbac S breathing filter have a low bacterial contamination rate. *Can J Anaesth* 2001;48:748–54.
90. Hiebert T, Miles J, Okeson GC. Contaminated aerosol recovery from pulmonary function testing equipment. *Am J Respir Crit Care Med* 1999;159:610–2.
91. Rutala DR, Rutala WA, Weber DJ, Thomann CA. Infection risks associated with spirometry. *Infect Control Hosp Epidemiol* 1991;12:89–92.
92. Ahmed J, Brutus A, D'Amato RF, Glatt AE. *Acinetobacter calcoaceticus anitratus* outbreak in the intensive care unit traced to a peak flow meter. *Am J Infect Control* 1994;22:319–21.
93. Griebble HG, Colton FR, Thomas MS, et al. Fine-particle humidifiers: source of *Pseudomonas aeruginosa* infections in a respiratory-disease unit. *N Engl J Med* 1970;282:531–3.
94. Smith PW, Massanari RM. Room humidifiers as the source of *Acinetobacter* infections. *JAMA* 1977;237:795–7.
95. CDC. Guidelines for environmental control in health-care facilities. *MMWR* 2003;52(No. RR-10).
96. Doebbeling BN, Pfaller MA, Houston AK, Wenzel RP. Removal of nosocomial pathogens from the contaminated glove: implications for glove reuse and handwashing. *Ann Intern Med* 1988;109:394–8.
97. LeClair JM, Freeman J, Sullivan BF, Crowley CM, Goldmann DA. Prevention of nosocomial respiratory syncytial virus infections through compliance with glove and gown isolation precautions. *N Engl J Med* 1987;317:329–34.
98. Patterson JE, Vecchio J, Pantelick EL, et al. Association of contaminated gloves with transmission of *Acinetobacter calcoaceticus* var. *anitratus* in an intensive care unit. *Am J Med* 1991;91:479–83.
99. Morar P, Makura Z, Jones A, et al. Topical antibiotics on tracheostoma prevents exogenous colonization and infection of lower airways in children. *Chest* 2000;117(2):513–8.
100. Combes P, Fauvage B, Oleyer C. Nosocomial pneumonia in mechanically ventilated patients: a prospective randomised evaluation of the Stericath closed suctioning system. *Intensive Care Med* 2000;26:878–82.
101. Deppe SA, Kelly JW, Thoi LL, et al. Incidence of colonization, nosocomial pneumonia, and mortality in critically ill patients using a Trach Care closed-suction system versus open-suction system: prospective, randomized study. *Crit Care Med* 1990;18:1389–93.
102. Johnson KL, Kearney PA, Johnson SB, Niblett JB, MacMillan NL, McClain RE. Closed versus open endotracheal suctioning: costs and physiologic consequences. *Crit Care Med* 1994;22:658–66.
103. Kollef MH, Prentice D, Shapiro SD, et al. Mechanical ventilation with or without daily changes of in-line suction catheters. *Am J Respir Crit Care Med* 1997;156:466–72.
104. CDC. Preventing pneumococcal disease among infants and young children. *MMWR* 2000;49(No. RR-9).
105. CDC. Prevention of pneumococcal disease: recommendations of the Advisory Committee on Immunization Practices (ACIP). *MMWR* 1997;46(No. RR-8).
106. CDC. Outbreak of pneumococcal pneumonia among unvaccinated residents of a nursing home—New Jersey, April 2001. *MMWR* 2001;50:707–10.
107. Shapiro ED, Clemens JD. A controlled evaluation of the protective efficacy of pneumococcal vaccine for patients at high risk of serious pneumococcal infections. *Ann Intern Med* 1984;101:325–30.
108. Williams WW, Hickson MA, Kane MA, Kendal AP, Spika JS, Hinman AR. Immunization policies and vaccine coverage among adults: the risk for missed opportunities. *Ann Intern Med* 1988;108:616–25.
109. Nichol KL, Grimm MB, Peterson DC. Immunizations in long-term care facilities: policies and practice. *J Am Geriatr Soc* 1996;44:349–55.
110. CDC. Use of standing orders programs to increase adult vaccination rates: recommendations of the Advisory Committee on Immunization Practices (ACIP). *MMWR* 2000;49(No. RR-1).
111. Centers for Medicare and Medicaid Services H. Medicare and Medicaid programs: conditions of participation: immunization standards for hospitals, long-term care facilities, and home health agencies: final rule with comment period. *Federal Register* 2002;67:61808–14.
112. Donta ST, Peduzzi P, Cross AS, et al. Immunoprophylaxis against *Klebsiella* and *Pseudomonas aeruginosa* infections. *J Infect Dis* 1996;174:537–43.

113. Gruson D, Hilbert G, Vargas F, et al. Impact of colony-stimulating factor therapy on clinical outcome and frequency rate of nosocomial infections in intensive care unit neutropenic patients. *Crit Care Med* 2000;28:3155–60.
114. Heard SO, Fink MP, Gamelli RL, et al. Effect of prophylactic administration of recombinant human granulocyte colony-stimulating factor (filgrastim) on the frequency of nosocomial infections in patients with acute traumatic brain injury or cerebral hemorrhage. *Crit Care Med* 1998;26:748–54.
115. Maher DW, Lieschke GJ, Green M, et al. Filgrastim in patients with chemotherapy-induced febrile neutropenia: a double-blind, placebo-controlled trial. *Ann Intern Med* 1994;121:492–501.
116. Mitchell PL, Morland B, Stevens MC, et al. Granulocyte colony-stimulating factor in established febrile neutropenia: a randomized study of pediatric patients. *J Clin Oncol* 1997;15:1163–70.
117. The Intravenous Immunoglobulin Collaborative Study Group. Prophylactic intravenous administration of standard immune globulin as compared with core-lipoplysaccharide immune globulin in patients at high risk of postsurgical infection. *N Engl J Med* 1992;327:234–40.
118. Houdijk APJ, Rijnsburger ER, Jansen J, et al. Randomised trial of glutamine-enriched enteral nutrition on infectious morbidity in patients with multiple trauma. *Lancet* 1998;352:772–6.
119. van der Hulst RRWJ, van Kreel BK, Von Meyenfeldt MF, et al. Glutamine and the preservation of gut integrity. *Lancet* 1993;341:1363–5.
120. Celis R, Torres A, Gatell JM, Almela M, Rodriguez-Riosin R, Agusti-Vidal A. Nosocomial pneumonia: a multivariate analysis of risk and prognosis. *Chest* 1988;93:318–24.
121. Craven DE, Kunches LM, Kilinsky V, Lichtenberg DA, Make BJ, McCabe WR. Risk factors for pneumonia and fatality in patients receiving continuous mechanical ventilation. *Am Rev Respir Dis* 1986;133:792–6.
122. Kingston GW, Phang PT, Leathley MJ. Increased incidence of nosocomial pneumonia in mechanically ventilated patients with subclinical aspiration. *Am J Surg* 1991;161:589–92.
123. Metheny NA, Eisenberg P, Spies M. Aspiration pneumonia in patients fed through nasogastric tubes. *Heart Lung* 1986;15:256–61.
124. Pingleton SK, Hinthorn DR, Liu C. Enteral nutrition in patients receiving mechanical ventilation: multiple sources of tracheal colonization include the stomach. *Am J Med* 1986;80:827–32.
125. Treloar DM, Stechmiller J. Pulmonary aspiration of tube-fed patients with artificial airways. *Heart Lung* 1984;13:667–71.
126. Brochard L, Mancebo J, Wysocki M, et al. Noninvasive ventilation for acute exacerbations of chronic obstructive pulmonary disease. *N Engl J Med* 1995;333:817–22.
127. Girou E, Schortgen F, Delcaux C, et al. Association of noninvasive ventilation with nosocomial infections and survival in critically ill patients. *JAMA* 2000;284:2361–7.
128. Carlucci A, Richard JC, Wysocki M, Lopage E, Brochard L. Noninvasive versus conventional mechanical ventilation: an epidemiologic survey. *Am J Respir Crit Care Med* 2001;163:874–80.
129. Keenan SP. Noninvasive positive pressure ventilation in acute respiratory failure. *JAMA* 2000;284:2376–8.
130. Nava S, Ambrosino N, Clini E, et al. Noninvasive mechanical ventilation in the weaning of patients with respiratory failure due to chronic obstructive pulmonary disease: a randomized, controlled trial. *Ann Intern Med* 1998;128:721–8.
131. Torres A, Gatell JM, Aznar E, et al. Re-intubation increases the risk of nosocomial pneumonia in patients needing mechanical ventilation. *Am J Respir Crit Care Med* 1995;152:137–41.
132. Holzapfel L, Chevret S, Madinier G, et al. Influence of long-term oro- or nasotracheal intubation on nosocomial maxillary sinusitis and pneumonia: results of a prospective, randomized clinical trial. *Crit Care Med* 1993;21:1132–8.
133. Rouby JJ, Laurent P, Gosnach M, et al. Risk factors and clinical relevance of nosocomial maxillary sinusitis in the critically ill. *Am J Respir Crit Care Med* 1994;150:776–83.
134. Kollef MH, Skubas NJ, Sundt TM. A randomized clinical trial of continuous aspiration of subglottic secretions in cardiac surgery patients. *Chest* 1999;116:1339–46.
135. Mahul P, Auboyer C, Jospe R, et al. Prevention of nosocomial pneumonia in intubated patients: respective role of mechanical subglottic drainage and stress ulcer prophylaxis. *Intensive Care Med* 1992;18:20–5.
136. Smulders K, van der Hoeven H, Weers-Pothoff I, Vanderbroucke-Grauls C. A randomized clinical trial of intermittent subglottic secretion drainage in patients receiving mechanical ventilation. *Chest* 2002;121:858–62.
137. Valles J, Artigas A, Rello J, et al. Continuous aspiration of subglottic secretions in preventing ventilator-associated pneumonia. *Ann Intern Med* 1995;122:179–86.
138. Drakulovic MB, Torres A, Bauer TT, Nicolas JM, Nogue S, Ferrer M. Supine body position as a risk factor for nosocomial pneumonia in mechanically ventilated patients: a randomised trial. *Lancet* 1999;354(9193):1851–58.
139. Orozco-Levi M, Torres A, Ferrer M, et al. Semirecumbent position protects from pulmonary aspiration but not completely from gastroesophageal reflux in mechanically ventilated patients. *Am J Respir Crit Care Med* 1995;152:1387–90.
140. Torres A, Serra-Batlles J, Ros E, et al. Pulmonary aspiration of gastric contents in patients receiving mechanical ventilation: the effect of body position. *Ann Intern Med* 1992;116:540–3.
141. Gharpure V, Meert KL, Sarnaik AP, Metheny NA. Indicators of postpyloric feeding tube placement in children. *Crit Care Med* 2000;28:2962–6.
142. Hand RW, Kempster M, Levy JH, Rogol PR, Spirn P. Inadvertent transbronchial insertion of narrow-bore feeding tubes into the pleural space. *JAMA* 1984;251:2396–7.
143. McClave SA, DeMeo MT, DeLegge MH, et al. North American summit on aspiration in the critically ill patient: consensus statement. *J Parenter Enter Nutr* 2002;26:80–5.
144. Ferrer M, Bauer TT, Torres A, Hernandez C, Pira C. Effect of nasogastric tube size on gastroesophageal reflux and microaspiration in intubated patients. *Ann Intern Med* 1999;130:991–4.
145. Bonten MJM, Gaillard CA, van der Hulst R, et al. Intermittent enteral feeding: the influence on respiratory and digestive tract colonization in mechanically ventilated intensive-care-unit patients. *Am J Respir Crit Care Med* 1996;154:394–9.
146. Jacobs S, Chang RW, Lee B, Bartlett FW. Continuous enteral feeding: a major cause of pneumonia among ventilated intensive care unit patients. *J Parenter Enter Nutr* 1990;14:353–6.
147. Lee B, Chang RWS, Jacobs S. Intermittent nasogastric feeding: a simple and effective method to reduce pneumonia among ventilated ICU patients. *Clin Intensive Care* 1990;1:100–2.
148. Skiest DJ, Khan N, Feld R, Metersky ML. The role of enteral feeding in gastric colonisation: a randomised controlled trial comparing continuous to intermittent enteral feeding in mechanically ventilated patients. *Clin Intensive Care* 1996;7:138–43.
149. Heyland DK, Drover JW, MacDonald S, Novak F, Lam M. Effect of postpyloric feeding on gastroesophageal regurgitation and pulmonary microaspiration. *Crit Care Med* 2001;29:1495–500.
150. Heyland DK, Drover JW, Dhaliwal R, Greenwood J. Optimizing the benefits and minimizing the risks of enteral nutrition in the critically ill: role of small bowel feeding. *J Parenter Enter Nutr* 2002;26:S51–7.

151. Kearns PJ, Chin D, Mueller L, Wallace K, Jensen WA, Kirsch CM. The incidence of ventilator-associated pneumonia and success in nutrient delivery with gastric versus small intestinal feeding: a randomized clinical trial. *Crit Care Med* 2000;28:1742-6.
152. Montecalvo M, Steger KA, Farber HW, et al. Nutritional outcome and pneumonia in critical care patients randomized to gastric versus jejunal tube feedings. *Crit Care Med* 1992;20:1377-87.
153. Montejo JC, Grau T, Acosta J, et al. Multicenter, prospective, randomized, single-blind study comparing the efficacy and gastrointestinal complications of early jejunal feeding with early gastric feeding in critically ill patients. *Crit Care Med* 2002;30:796-800.
154. Spain DA, DeWeese RC, Reynolds MA, Richardson JD. Transpyloric passage of feeding tubes in patients with head injuries does not decrease complications. *J Trauma* 1995;39:1000-2.
155. Strong RM, Condon SC, Solinger MR, Namihas BN, Ito-Wong LA, Leuty JE. Equal aspiration rates from postpylorus and intragastric-placed small-bore nasogastric feeding tubes: a randomized, prospective study. *J Parent Enter Nutr* 1992;16:59-63.
156. Schleder B, Stott K, Lloyd RC. The effect of a comprehensive oral care protocol on patients at risk for ventilator-associated pneumonia. *J Advocate Health Care* 2002;4:27-30.
157. Yoneyama T, Yoshida M, Ohru T, et al. Oral care reduces pneumonia in older patients in nursing homes. *J Am Geriatr Soc* 2002;50:430-3.
158. DeRiso AJII, Ladowski JS, Dillon TA, Justice JW, Peterson AC. Chlorhexidine gluconate 0.12% oral rinse reduces the incidence of total nosocomial respiratory infections and nonprophylactic antibiotic use in patients undergoing heart surgery. *Chest* 1996;109:1556-61.
159. Bergmans D, Bonten M, Gaillard C, et al. Prevention of ventilator-associated pneumonia by oral decontamination: a prospective, randomized, double-blind, placebo-controlled study. *Am J Respir Crit Care Med* 2001;164:382-8.
160. Bonten MJM, Gaillard CA, van der Geest S, et al. The role of intragastric acidity and stress ulcer prophylaxis on colonization and infection in mechanically ventilated patients: a stratified, randomized, double-blind study of sucralfate versus antacids. *Am J Respir Crit Care Med* 1995;152:1825-34.
161. Cook D, Guyatt G, Marshall J, et al. A comparison of sucralfate and ranitidine for the prevention of upper gastrointestinal bleeding in patients requiring mechanical ventilation. *N Engl J Med* 1998;338:791-7.
162. Cook DJ, Reeve BK, Guyatt GH, et al. Stress ulcer prophylaxis in critically ill patients: resolving discordant meta-analyses. *JAMA* 1996;275:308-14.
163. Messori A, Trippoli SI, Vaiani M, Gorini M, Corrado A. Bleeding and pneumonia in intensive care patients given ranitidine and sucralfate for prevention of stress ulcer: meta-analysis of randomised controlled trials. *BMJ* 2000;321:1103-6.
164. Simms HH, DeMaria E, McDonald L, Peterson D, Robinson A, Burchard KW. Role of gastric colonization in the development of pneumonia in critically ill trauma patients: results of a prospective randomized trial. *J Trauma* 1991;31:531-6.
165. Thomason MH, Payseur ES, Hakenewerth AM, et al. Nosocomial pneumonia in ventilated trauma patients during stress ulcer prophylaxis with sucralfate, antacid, and ranitidine. *J Trauma-Injury Infect Crit Care* 1996;41:503-8.
166. Tryba M. Risk of acute stress bleeding and nosocomial pneumonia in ventilated intensive care unit patients: sucralfate versus antacids. *Am J Med* 1987;83:117-24.
167. Yildizdas D, Yapicioglu H, Yilmaz HL. Occurrence of ventilator-associated pneumonia in mechanically ventilated pediatric intensive care patients during stress ulcer prophylaxis with sucralfate, ranitidine, and omeprazole. *J Crit Care* 2002;17:240-5.
168. Abele-Horn M, Dauber A, Bauernfeind A, et al. Decrease in nosocomial pneumonia in ventilated patients by selective oropharyngeal decontamination: a prospective, blinded, randomized trial of the effect of a novel regimen. *Intensive Care Med* 1997;23:187-95.
169. D'Amico R, Pifferi S, Leonetti C, Torri V, Tinazzi A, Liberati A. Effectiveness of antibiotic prophylaxis in critically ill patients: systemic review of randomised controlled trials. *BMJ* 1998;316:1275-85.
170. Langlois-Karaga A, Bues-Charbit M, Davignon A, et al. Selective digestive decontamination in multiple trauma patients: cost and efficacy. *Pharmacy World and Science* 1995;17:12-6.
171. Nathens AB, Marshall JC. Selective decontamination of the digestive tract in surgical patients: a systematic review of the evidence. *Arch Surg* 1999;134:170-6.
172. Quinio B, Albanese J, Bues-Charbit M, Viviani X, Martin C. Selective decontamination of the digestive tract in multiple trauma patients: a prospective double-blind, randomized, placebo-controlled study. *Chest* 1996;109:765-72.
173. Stoutenbeek CP, Van Saene HKF, Miranda DR, Zandstra DF. The effect of selective decontamination of the digestive tract on colonisation and infection rate in multiple trauma patients. *Intensive Care Med* 1984;10:185-92.
174. Unertl K, Ruckdeschel G, Selbmann HK, et al. Prevention of colonization and respiratory infections in long-term ventilated patients by local antimicrobial prophylaxis. *Intensive Care Med* 1987;13:106-13.
175. Kerver JH, Rommes JH, Mevissen-Verhage EAE, et al. Prevention of colonization and infection in critically ill patients: a prospective randomized study. *Crit Care Med* 1988;16:1087-93.
176. Ledingham IM, Alcock SR, Eastaway AT, McDonald JC, McKay IC, Ramsay G. Triple regimen of selective decontamination of the digestive tract, systemic cefotaxime, and microbiological surveillance for prevention of acquired infection in intensive care. *Lancet* 1988;1:785-90.
177. Brun-Buisson C, Legrand P, Rauss A, et al. Intestinal decontamination for control of nosocomial multiresistant gram-negative bacilli: study of an outbreak in an intensive care unit. *Ann Intern Med* 1989;110:873-81.
178. Ulrich S, Harinck-de Weerd JE, Bakker NC, Jacz K, Doornbos L, de Ridder VA. Selective decontamination of the digestive tract with norfloxacin in the prevention of ICU-acquired infections: a prospective randomized study. *Intensive Care Med* 1989;15:424-31.
179. Flaherty J, Nathan C, Kabins SA, Weinstein RA. Pilot trial of selective decontamination for prevention of bacterial infection in an intensive care unit. *J Infect Dis* 1990;162:1393-7.
180. Godard J, Guillaume C, Reverdy ME, et al. Intestinal decontamination in a polyvalent ICU: a double-blind study. *Intensive Care Med* 1990;16:307-11.
181. McClelland P, Murray AE, Williams PS, et al. Reducing sepsis in severe combined acute renal and respiratory failure by selective decontamination of the digestive tract. *Crit Care Med* 1990;18:935-9.
182. Rodriguez-Roldan JM, Altuna-Cuesta A, Lopez A, et al. Prevention of nosocomial lung infection in ventilated patients: use of an antimicrobial pharyngeal non-absorbable paste. *Crit Care Med* 1990;18:1239-42.
183. Tetteroo GWM, Wagenvoort JHT, Castelein A, Tilanus HW, Ince C, Bruining HA. Selective decontamination to reduce gram-negative colonisation and infections after oesophageal resection. *Lancet* 1990;335:704-7.
184. Aerdt SJA, van Daelen R, Clasener HAL, Festen J, Van Lier HJJ, Vollaard EJ. Antibiotic prophylaxis of respiratory tract infection in mechanically ventilated patients: a prospective, blinded, randomized trial of the effect of a novel regimen. *Chest* 1991;100:783-91.

185. Blair P, Rowlands BJ, Lowry K, Webb H, Armstrong P, Smilie J. Selective decontamination of the digestive tract: a stratified, randomized, prospective study in a mixed intensive care unit. *Surgery* 1991;110:303–10.
186. Fox MA, Peterson S, Fabri BM, Van Saene HKF, Williets T. Selective decontamination of the digestive tract in cardiac surgical patients. *Crit Care Med* 1991;19:1486–90.
187. Hartenauer U, Thulig B, Diemer W, et al. Effect of selective flora suppression on colonization, infection and mortality in critically ill patients: a one-year, prospective, consecutive study. *Crit Care Med* 1991;19:463–73.
188. Pugin J, Auckenthaler R, Lew DP, Suter PM. Oropharyngeal decontamination decreases incidence of ventilator-associated pneumonia: a randomized, placebo-controlled, double-blind clinical trial. *JAMA* 1991;265:2704–10.
189. Vandembroucke-Grauls CMJE, Vandembroucke JP. Effect of selective decontamination of the digestive tract on respiratory tract infections and mortality in the intensive care unit. *Lancet* 1991;338:859–62.
190. Cockerill FR, Muller SM, Anhalt JP, et al. Prevention of infection on critically ill patients by selective decontamination of the digestive tract. *Ann Intern Med* 1992;117:545–53.
191. Gastinne H, Wolff M, Delatour F, Faurisson F, Chevret S. A controlled trial in intensive care units of selective decontamination of the digestive tract with nonabsorbable antibiotics. *N Engl J Med* 1992;326:594–9.
192. Hammond JMJ, Potgieter PD, Saunders GL, Forder AA. Double-blind study of selective decontamination of the digestive tract in intensive care. *Lancet* 1992;340:5–9.
193. Rocha LA, Martin MJ, Pita S, et al. Prevention of nosocomial infection in critically ill patients by selective decontamination of the digestive tract. A randomized, double-blind, placebo-controlled study. *Intensive Care Med* 1992;18:398–404.
194. Winter R, Humphreys H, Pick A, MacGowan P, Willatts SM, Speller DCE. A controlled trial of selective decontamination of the digestive tract in intensive care and its effect on nosocomial infection. *J Antimicrob Chemother* 1992;30:73–87.
195. Korinek AM, Laisne MJ, Nicolas MH, Raskine L, Deroin V, Sanson-Lepors MJ. Selective decontamination of the digestive tract in neurosurgical intensive care unit patients: a double-blind, randomized, placebo-controlled study. *Crit Care Med* 1993;21:1466–73.
196. Selective Decontamination of the Digestive Tract Trialists' Collaborative Group. Meta-analysis of randomised controlled trials of selective decontamination of the digestive tract. *BMJ* 1993;307:525–32.
197. Ferrer M, Torres A, Gonzalez J, et al. Utility of selective digestive decontamination in mechanically ventilated patients. *Ann Intern Med* 1994;120:389–95.
198. Nau R, Ruchel R, Mergerian H, Wegener U, Winkelmann T, Prange HW. Emergence of antibiotic-resistant bacteria during selective decontamination of the digestive tract. *J Antimicrob Chemother* 1990;25:881–3.
199. Sanchez Garcia M, Cambronero Galache JA, Lopez Diaz J, et al. Effectiveness and cost of selective decontamination of the digestive tract in critically ill intubated patients. A randomized, double-blind, placebo-controlled, multicenter trial. *Am J Respir Crit Care Med* 1998;158:908–16.
200. Krueger WA, Lenhart FP, Neeser G, Ruckdeschel G, Schreckhase H, Eissner HJ et al. Influence of combined intravenous and topical antibiotic prophylaxis on the incidences of infections, organ dysfunction, and mortality in critically ill surgical patients: a prospective, stratified, randomized, double-blind, placebo-controlled clinical trial. *Am J Respir Crit Care Med* 2002;166:1029–37.
201. Heyland DK, Bradley C, Mandell LA. Effect of acidified enteral feedings on gastric colonization in the critically ill patient. *Crit Care Med* 1992;20:1388–94.
202. Heyland DK, Cook DJ, Schoenfeld PS, Frietag A, Varon J, Wood G. The effect of acidified enteral feeds on gastric colonization in critically ill patients: results of a multicenter randomized trial. *Crit Care Med* 1999;27:2399–406.
203. Arozullah AM, Khuri SF, Henderson WG, Daley J, Participants in the National Veterans Affairs Surgical Quality Improvement Program: development and validation of a multifactorial risk index for predicting postoperative pneumonia after major noncardiac surgery. *Ann Intern Med* 2001;135:847–57.
204. Brooks-Brunn JA. Predictors of postoperative pulmonary complications following abdominal surgery. *Chest* 1997;111:564–71.
205. Chumillas S, Ponce JL, Delgado F, Viciano V, Mateu M. Prevention of postoperative pulmonary complications through respiratory rehabilitation: a controlled clinical study. *Arch Phys Med Rehab* 1998;79:5–9.
206. Thomas JA, McIntosh JM. Are incentive spirometry, intermittent positive pressure breathing, and deep breathing exercises effective in the prevention of postoperative pulmonary complications after upper abdominal surgery? A systematic overview and meta-analysis. *Physical Therapy* 1994;74:3–10.
207. Hall JC, Tarala RA, Tapper J, Hall JL. Prevention of respiratory complications after abdominal surgery: a randomised clinical trial. *BMJ* 1996;312:148–52.
208. Sirvent JM, Torres A, El-ebiary M, Castro P, de Batlle J, Bonet A. Protective effect of intravenously administered cefuroxime against nosocomial pneumonia in patients with structural coma. *Am J Respir Crit Care Med* 1997;155:1729–34.
209. Gruson D, Hilbert G, Vargas F, et al. Rotation and restricted use of antibiotics in a medical intensive care unit: impact on the incidence of ventilator-associated pneumonia caused by antibiotic-resistant gram-negative bacteria. *Am J Respir Crit Care Med* 2000;162:837–43.
210. Kollef MH, Vlasnik J, Sharpless L, Pasque C, Murphy D, Fraser V. Scheduled change of antibiotic classes: a strategy to decrease the incidence of ventilator-associated pneumonia. *Am J Respir Crit Care Med* 1997;156:1040–8.
211. deBoisblanc BP, Castro M, Everret B, Grender J, Walker CD, Summer WR. Effect of air-supported, continuous, postural oscillation on the risk of early ICU pneumonia in nontraumatic critical illness. *Chest* 1993;103:1543–7.
212. Fink MP, Helmsmoortel CM, Stein KL, Lee PC, Cohn SM. The efficacy of an oscillating bed in the prevention of lower respiratory tract infection in critically ill victims of blunt trauma: a prospective study. *Chest* 1990;97:132–7.
213. Gentilello L, Thompson DA, Tonnesen AS, et al. Effect of a rotating bed on the incidence of pulmonary complications in critically ill patients. *Crit Care Med* 1988;16:783–6.
214. Kirschenbaum L, Azzi E, Sfeir T, Tietjen P, Astiz M. Effect of continuous lateral rotational therapy on the prevalence of ventilator-associated pneumonia in patients requiring long term ventilatory care. *Crit Care Med* 2002;30:1983–6.
215. Summer WR, Curry P, Haponik EF, Nelson S, Elston R. Continuous mechanical turning of intensive care unit patients shortens length of stay in some diagnostic-related groups. *J Crit Care* 1989;4:45–53.
216. Whiteman K, Nachtmann L, Kramer D, Sereika S, Bierman M. Effects of continuous lateral rotation therapy on pulmonary complications in liver transplant patients. *Am J Crit Care* 1995;4:133–9.